

# BIOENG-320

# Synthetic Biology

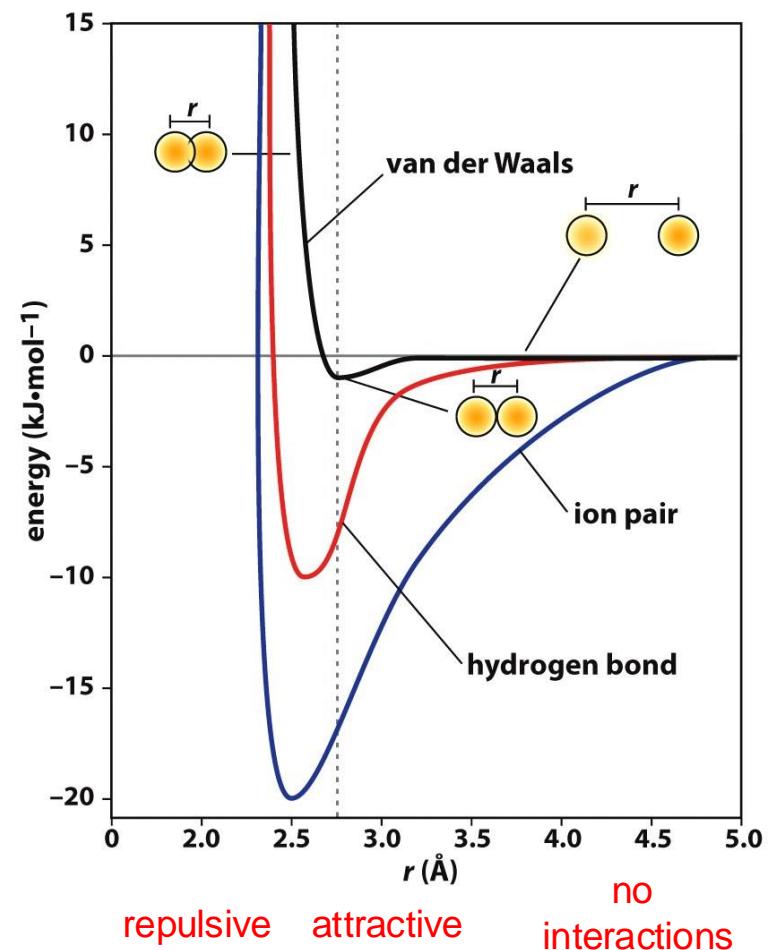
Protein design Lecture 3  
March 10, 2025

Design of protein binding

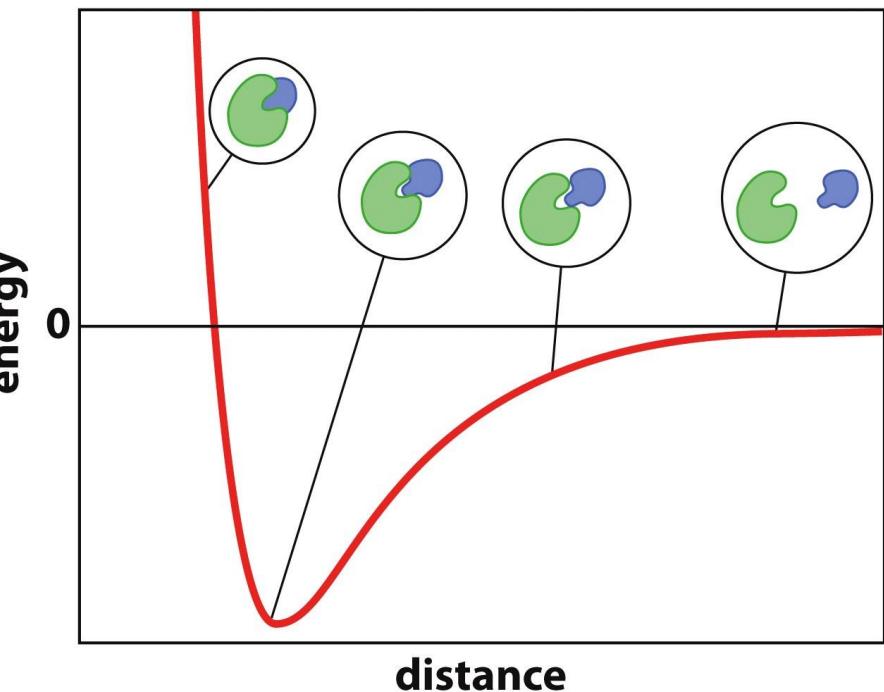
Patrick Barth  
EPFL

# Protein Design – the binding problem

Atomic interactions

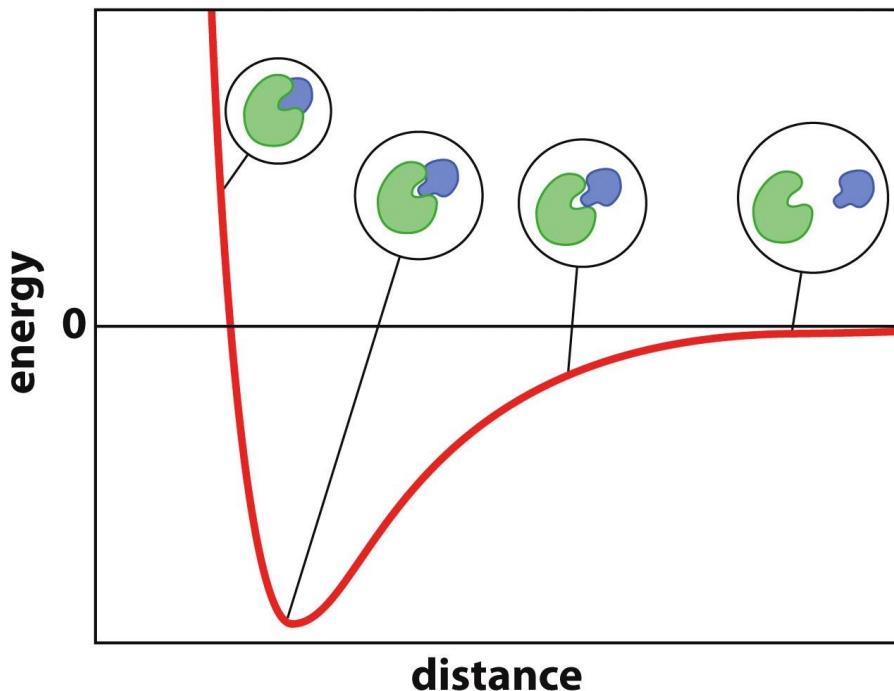


Protein-ligand interactions



# Protein Design – the binding problem

## Protein-ligand interactions



Similar forces than during protein folding:

1. Desolvation of 2 protein surfaces
2. Creation of hydrophobic and polar interactions at the binding interface

Balance between 1 and 2 will dictate the strength of binding

→ Protein surface complementarity and surface areas are key!

Buried cavity

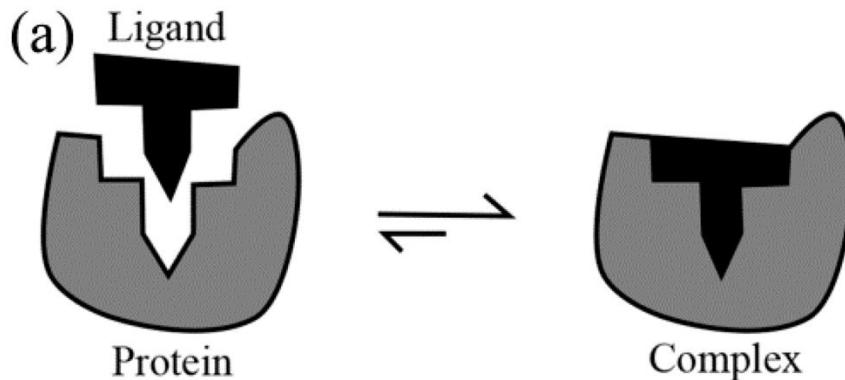


Flat binding surface



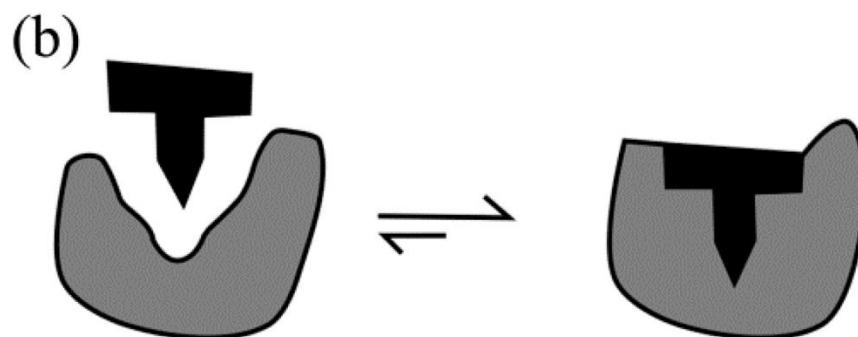
# Protein binding – a conformational flexibility problem

Lock and key



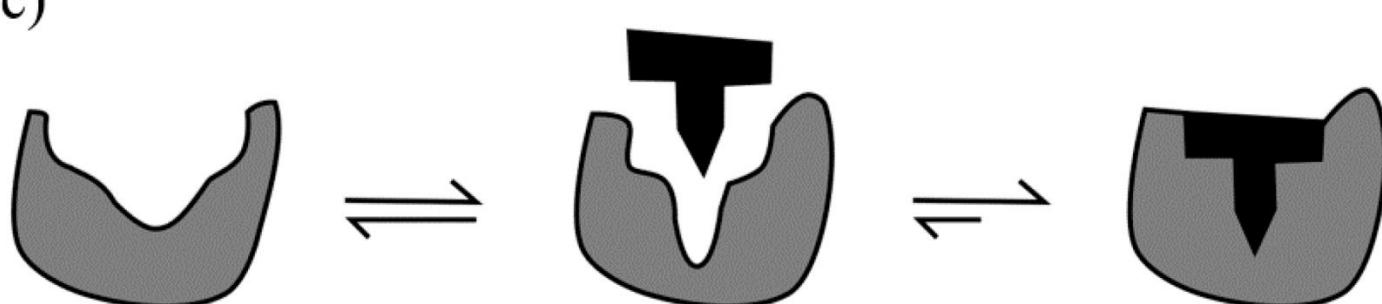
Rigid body  
conformational  
degrees  
of freedom

Induced fit



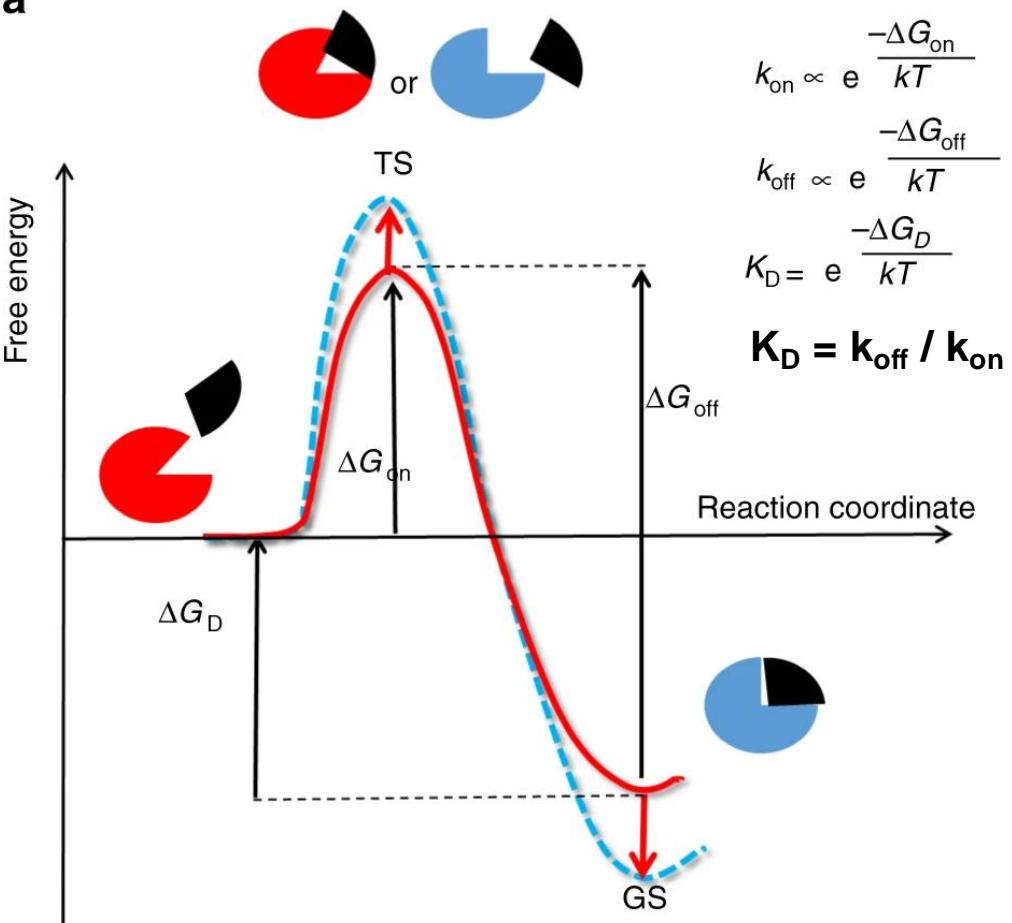
Rigid body + intraprotein  
conformational  
degrees  
of freedom

Conformational  
selection

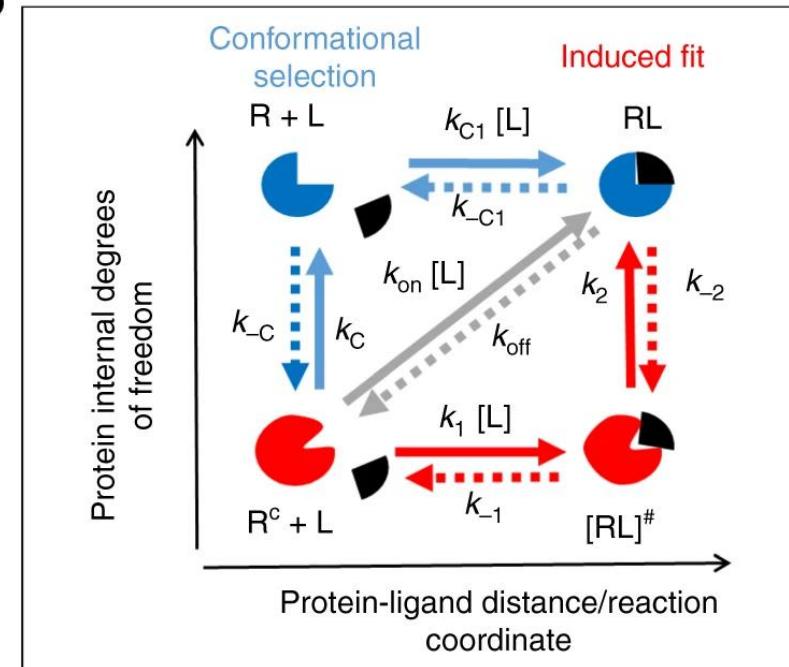


# Protein binding – a kinetic problem

a



b

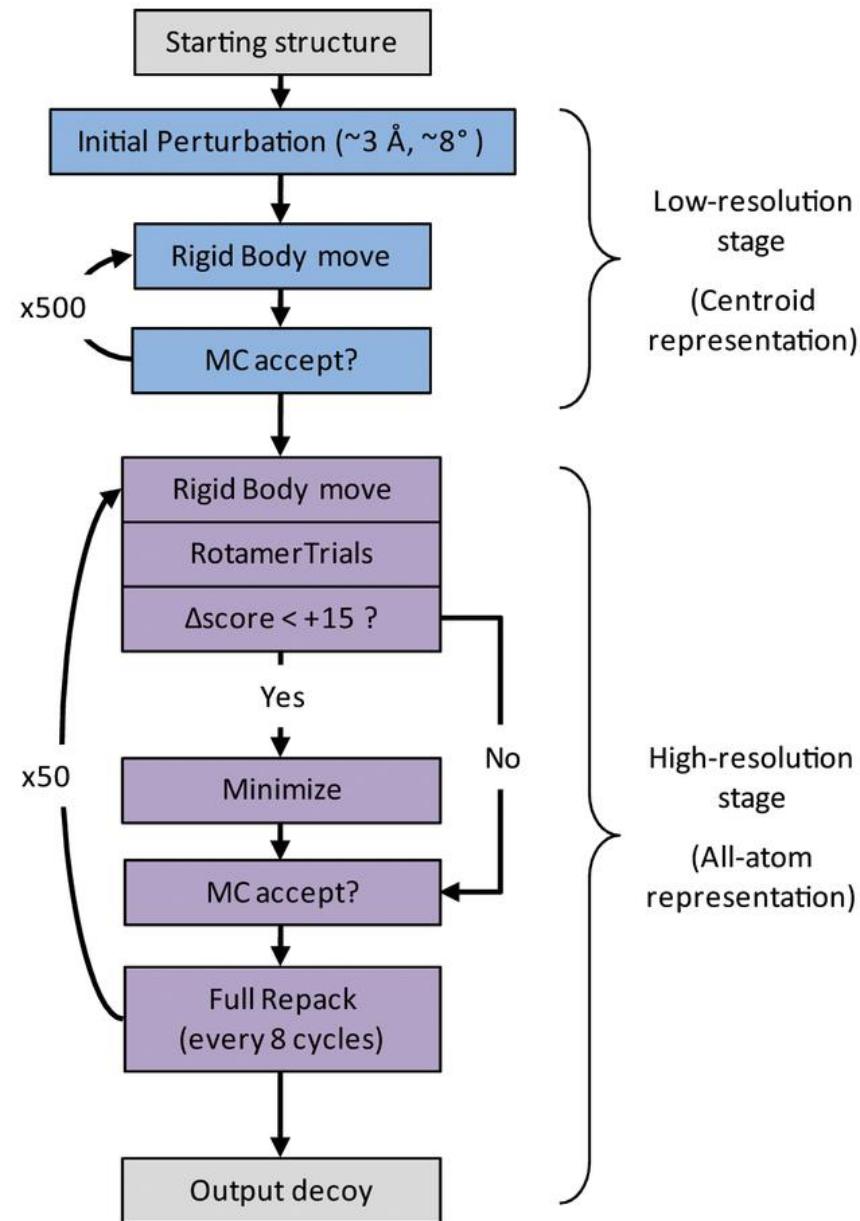
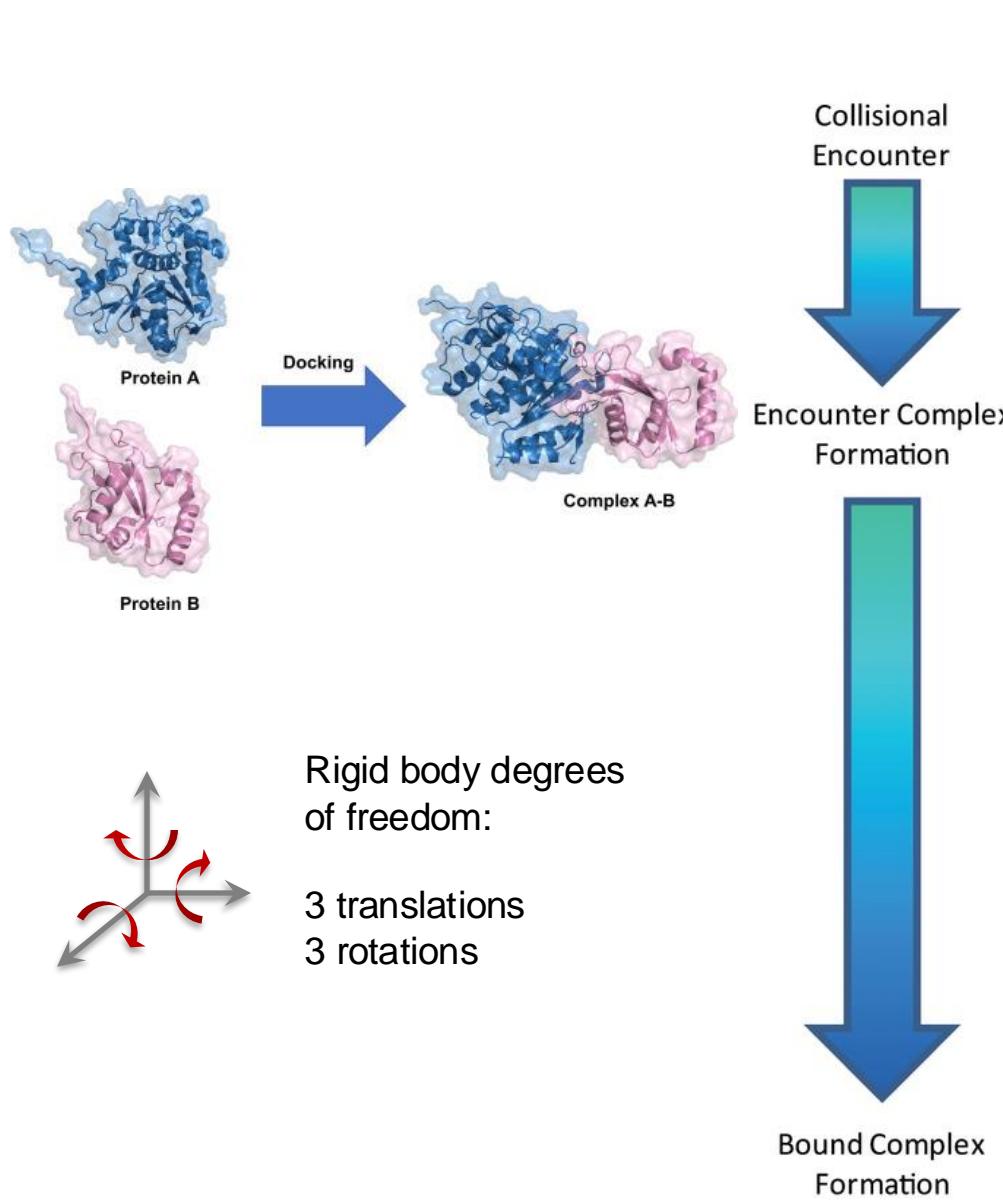


Conformational selection

Induced fit

Experimentally observable

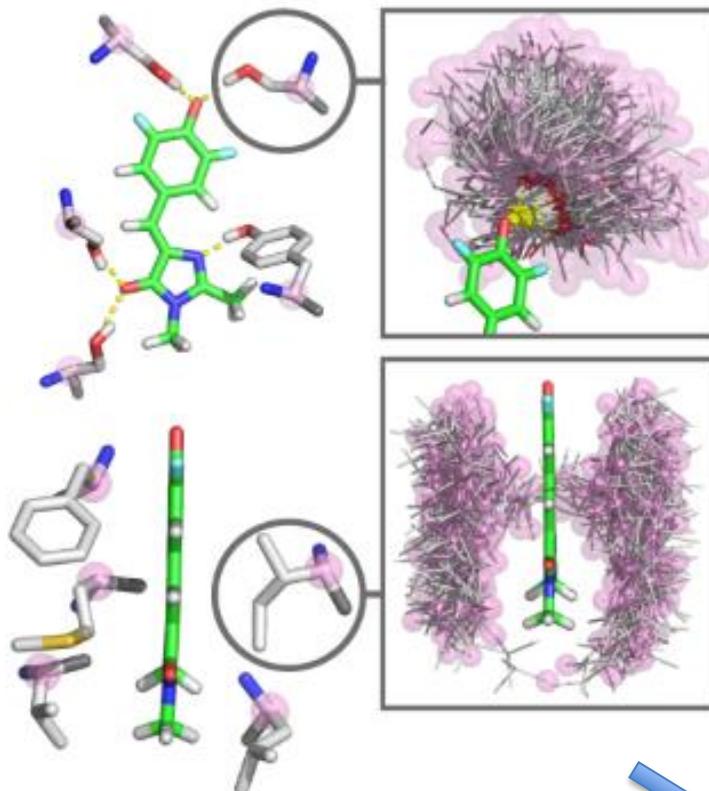
# Protein docking: prediction of binding complexes



# De novo design of ligand binding

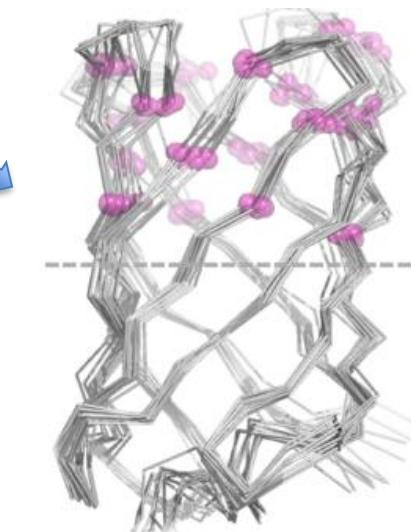
Step 1:  
Pre-defined optimal  
binding interactions  
between ligand &  
residue side-chains

●  $C\alpha$  atom



Step 2:  
Inverse rotamer  
generation from  
ligand-side-chain  
contacts

Step 3:  
Matching and grafting of ligand-binding  
residues onto a protein scaffold



# Protein Design – Examples overview

1. De novo protein functional fold

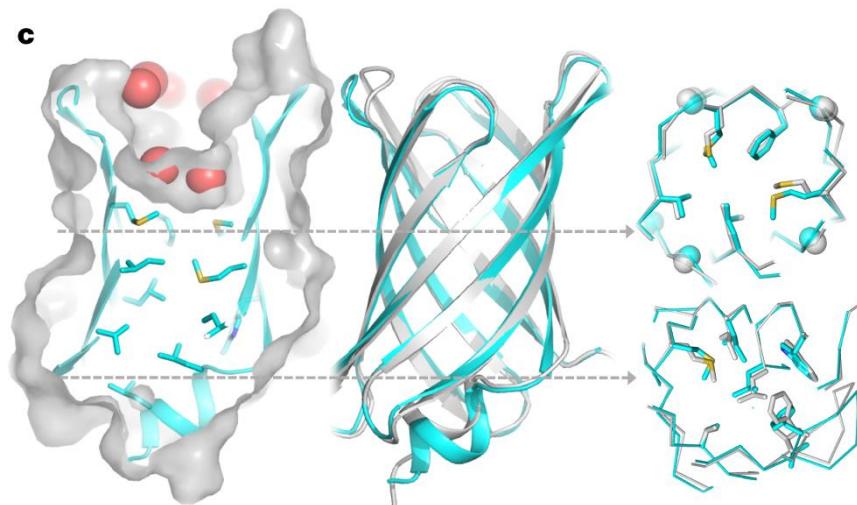
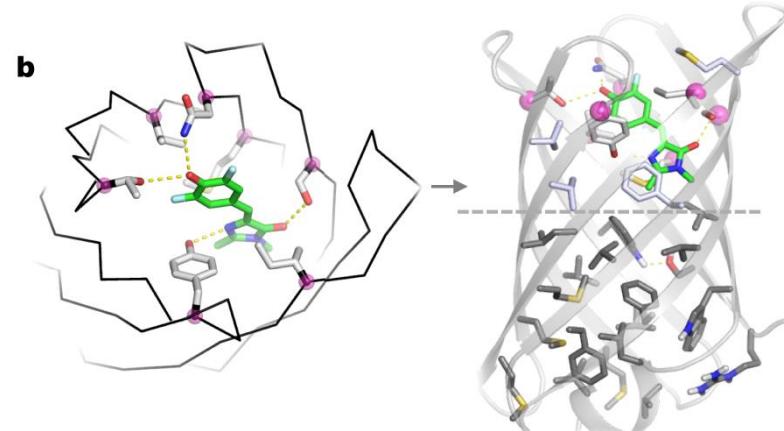
2. Enzyme

3. Ligand biosensor design

# Design of new functional folds: fluorescence activating beta barrel

The beta barrel  
scaffold

Ideal for ligand binding



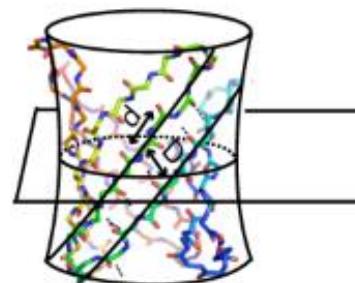
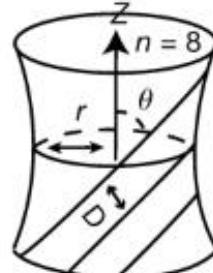
# Design of new functional folds: fluorescence activating beta barrel

3D ideal  
geometry

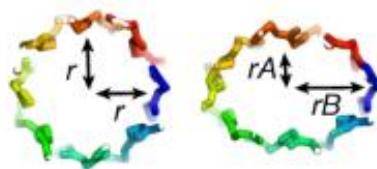


*parametrization  
using an equation for  
an elliptic hyperboloid  
of revolution*

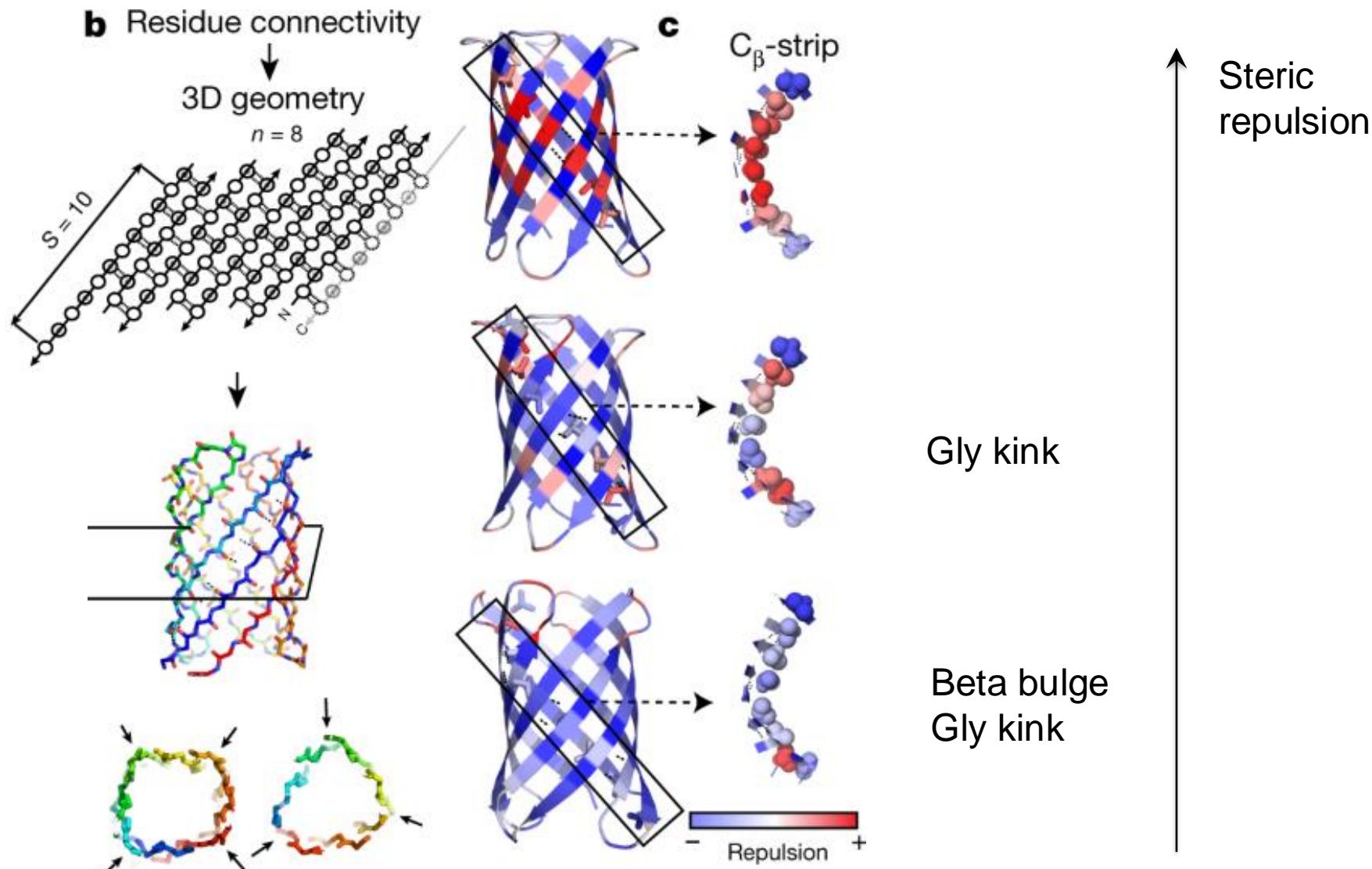
Residue connectivity



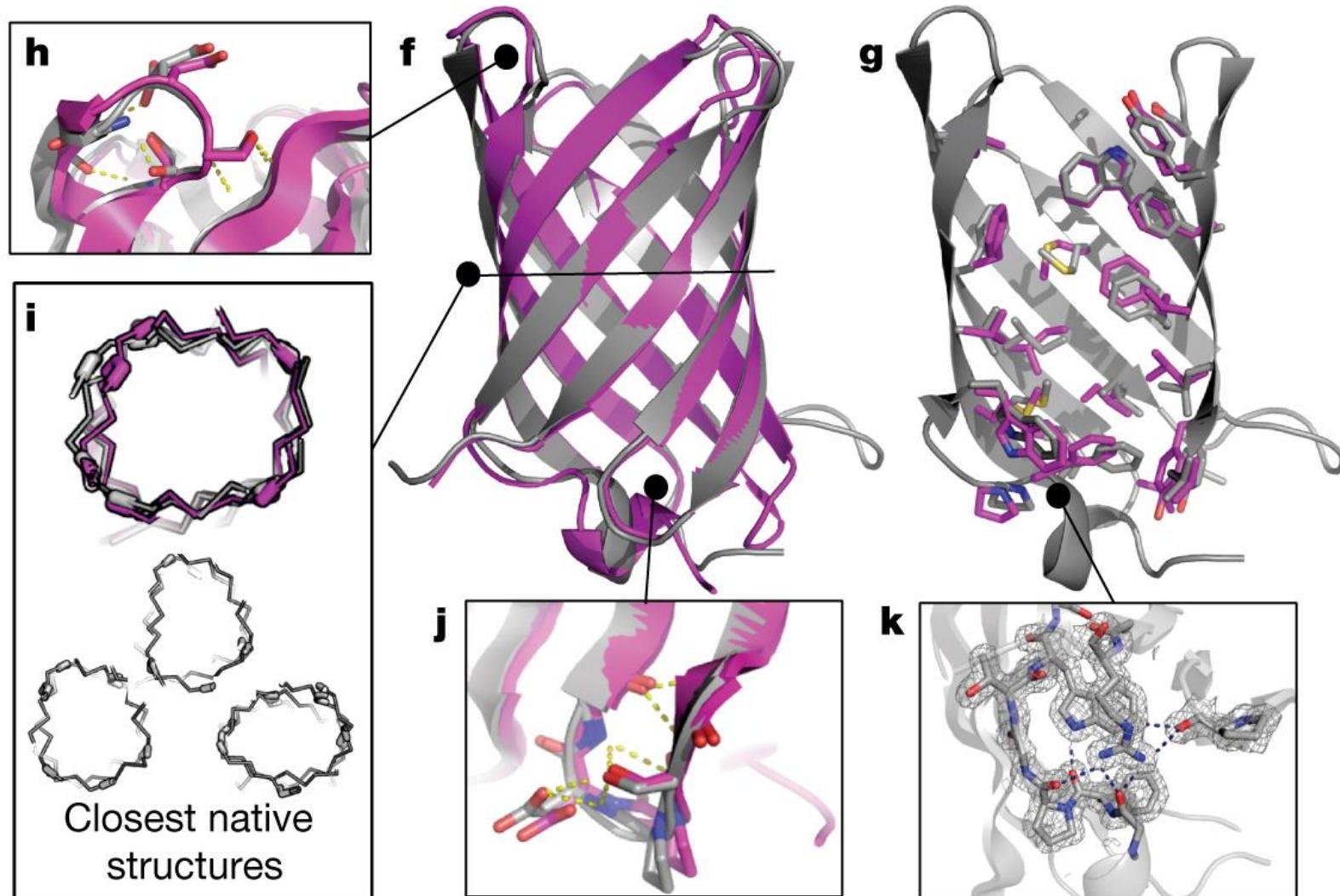
No  
folded  
designs !



# Design of new functional folds: fluorescence activating beta barrel

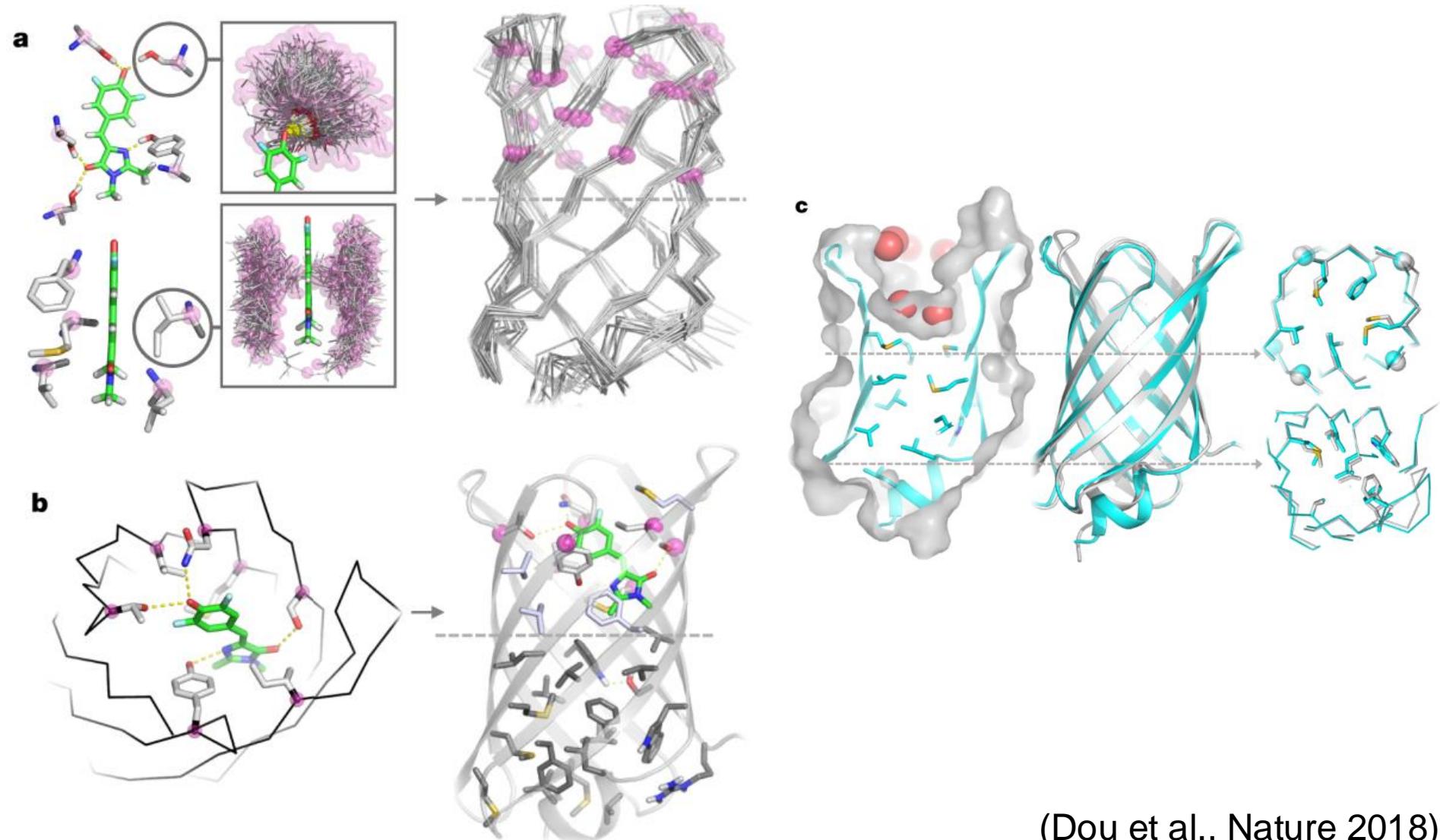


# Design of new functional folds: fluorescence activating beta barrel



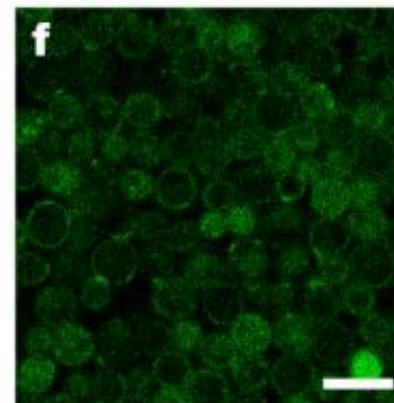
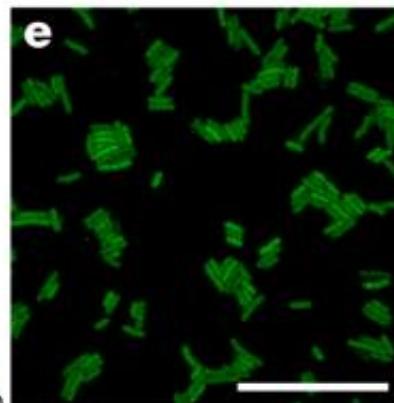
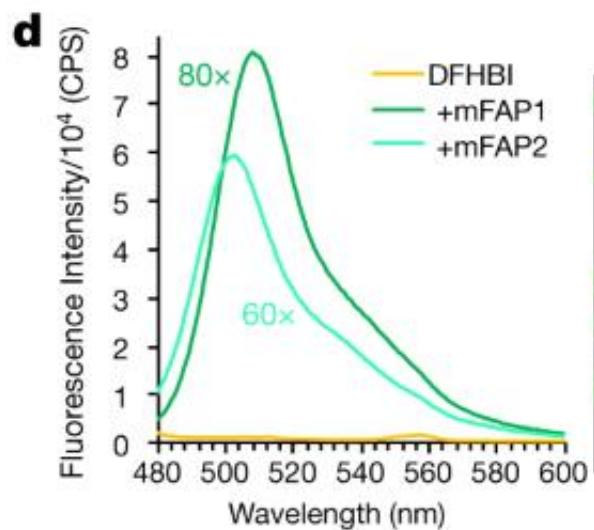
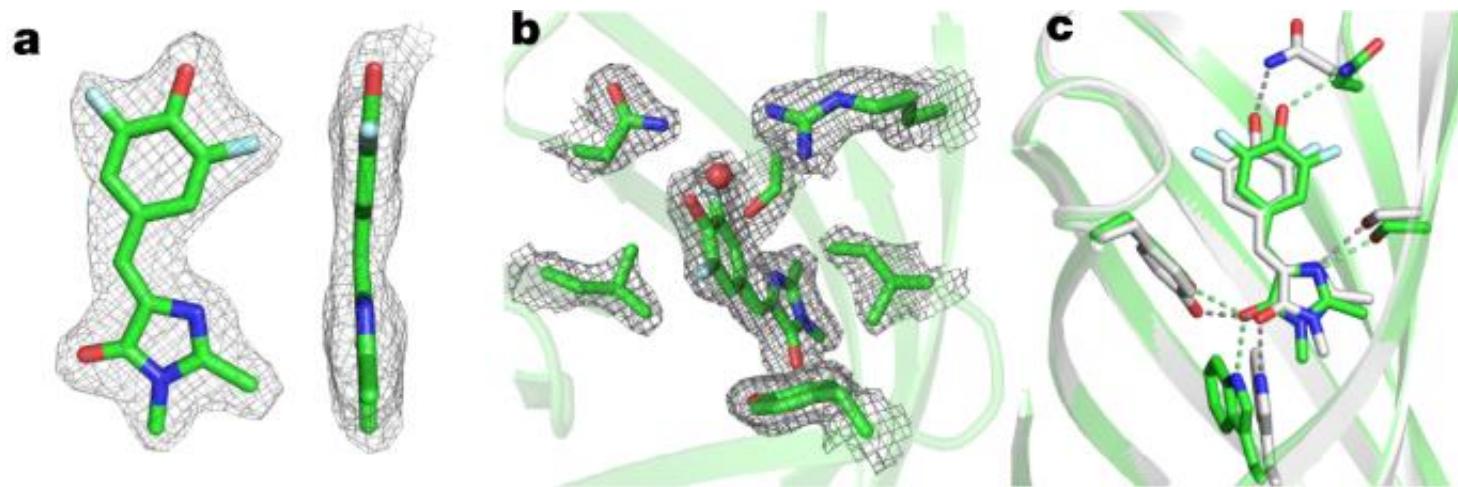
# Design of new functional folds

Barrel design with ligand binding cavity



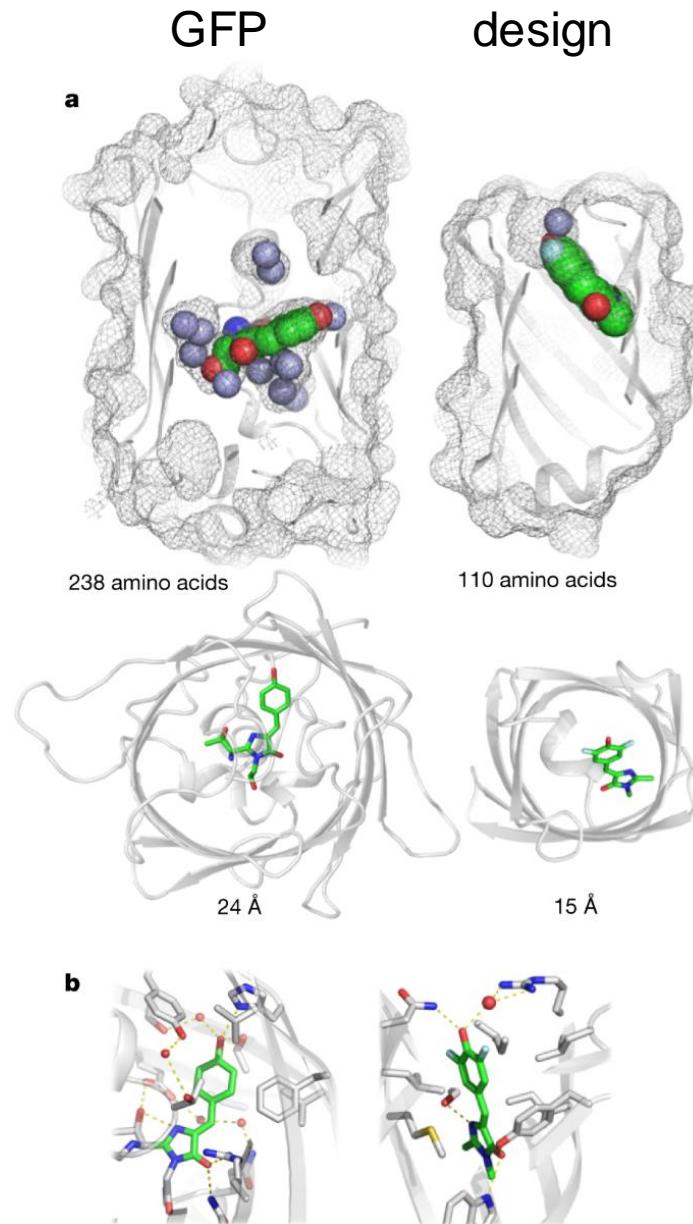
(Dou et al., Nature 2018)

# Design of new functional folds



# Design of new functional folds

## Comparison with naturally evolved GFP



# Take home messages

1. De novo design of beta barrels is challenging
2. Are the Gly kink bulge rules universal and applicable for all barrels?
3. Precise ligand binding is possible when starting from a hyperstable and rigid scaffold: destabilizing ligand binding cavity carved into a large hydrophobic stabilizing core

# Protein Design – Examples overview

1. De novo protein functional fold

2. Enzyme

3. Ligand biosensor design

# Design of a novel enzyme

- Goal: design **artificial enzymes** that catalyze unnatural reactions
- Enzymes:
  - lower the activation barrier, by
  - stabilizing transition state
  - shielding reactants

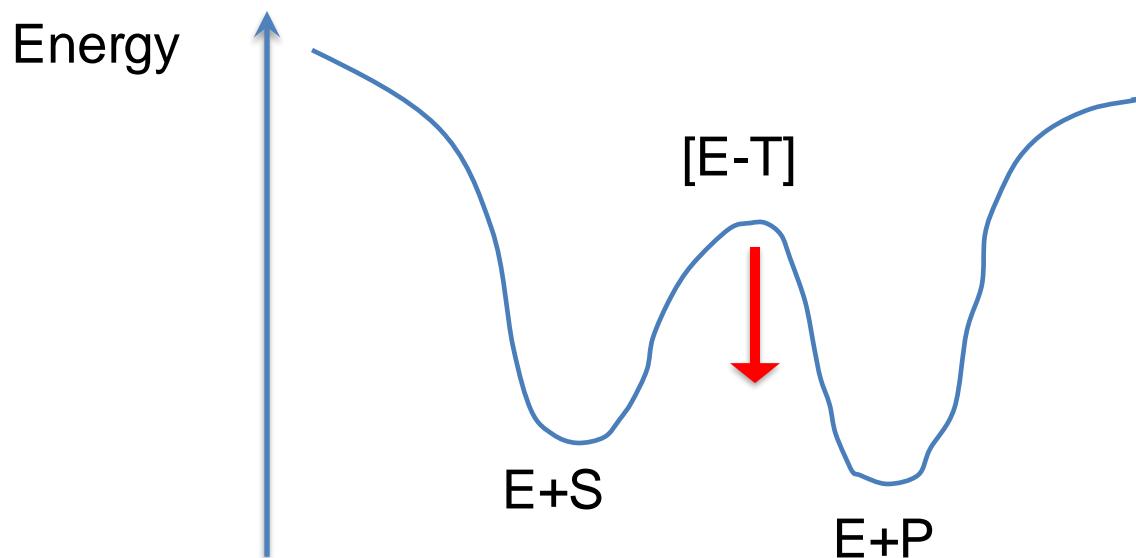


*Roethlisberger et al. 2008; Liang et al., 2008*

# Design of a novel enzyme

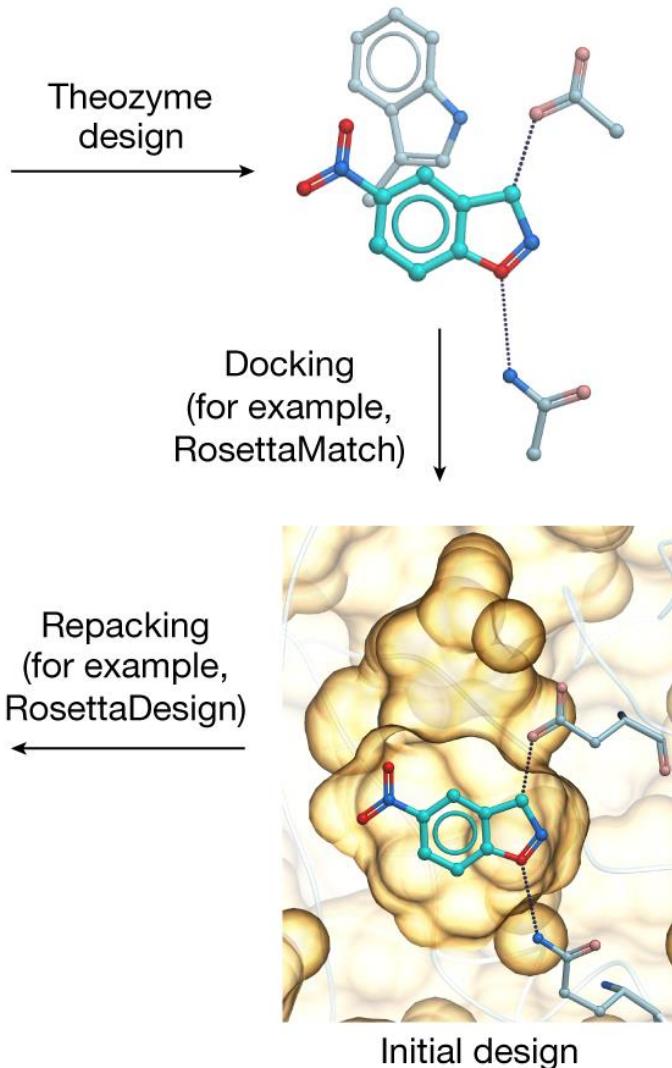
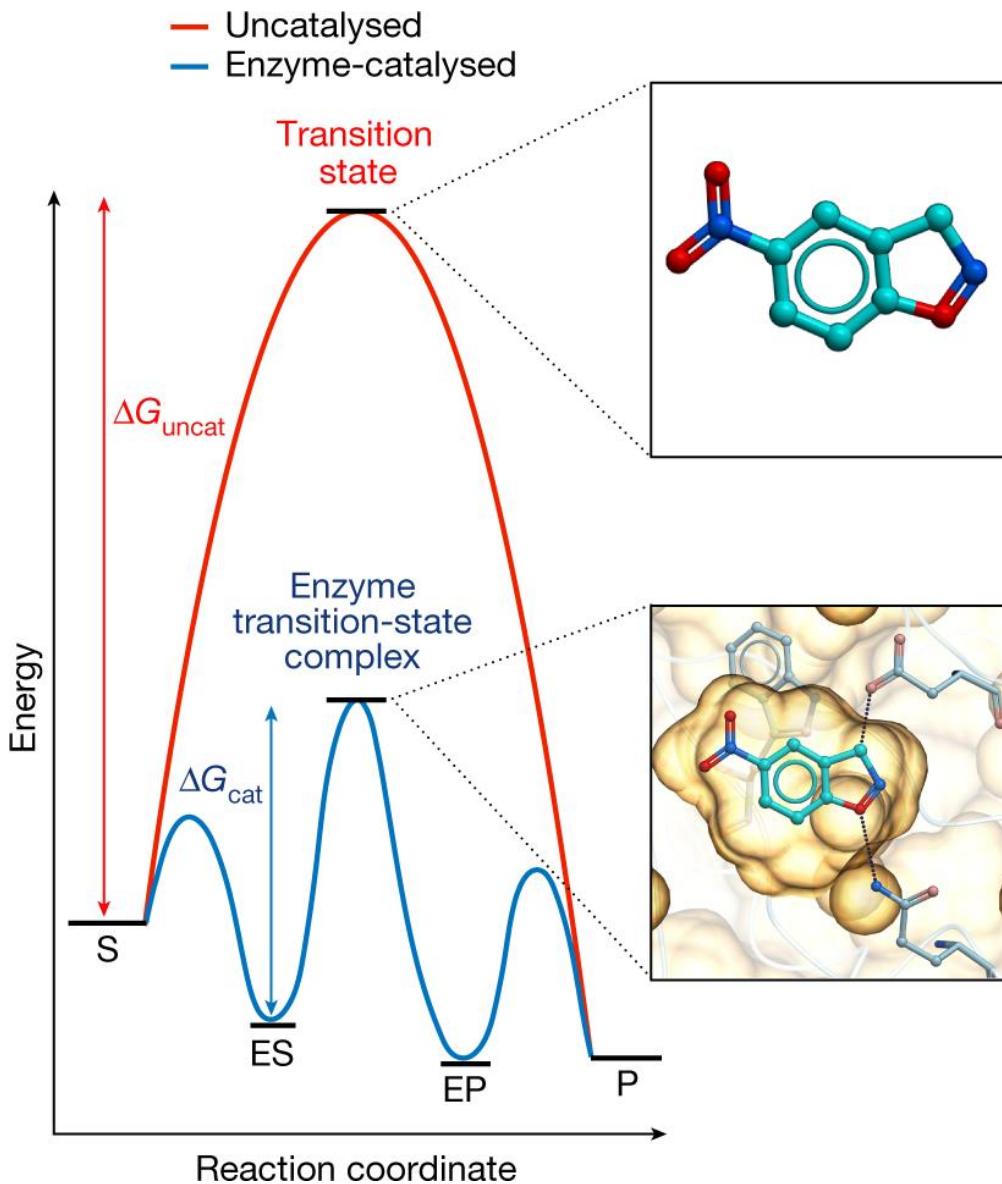
*Roethlisberger et al. 2008; Liang et al., 2008*

Enzyme + Substrate  $\rightarrow$  [ E-T ]  $\rightarrow$  Enzyme + Product



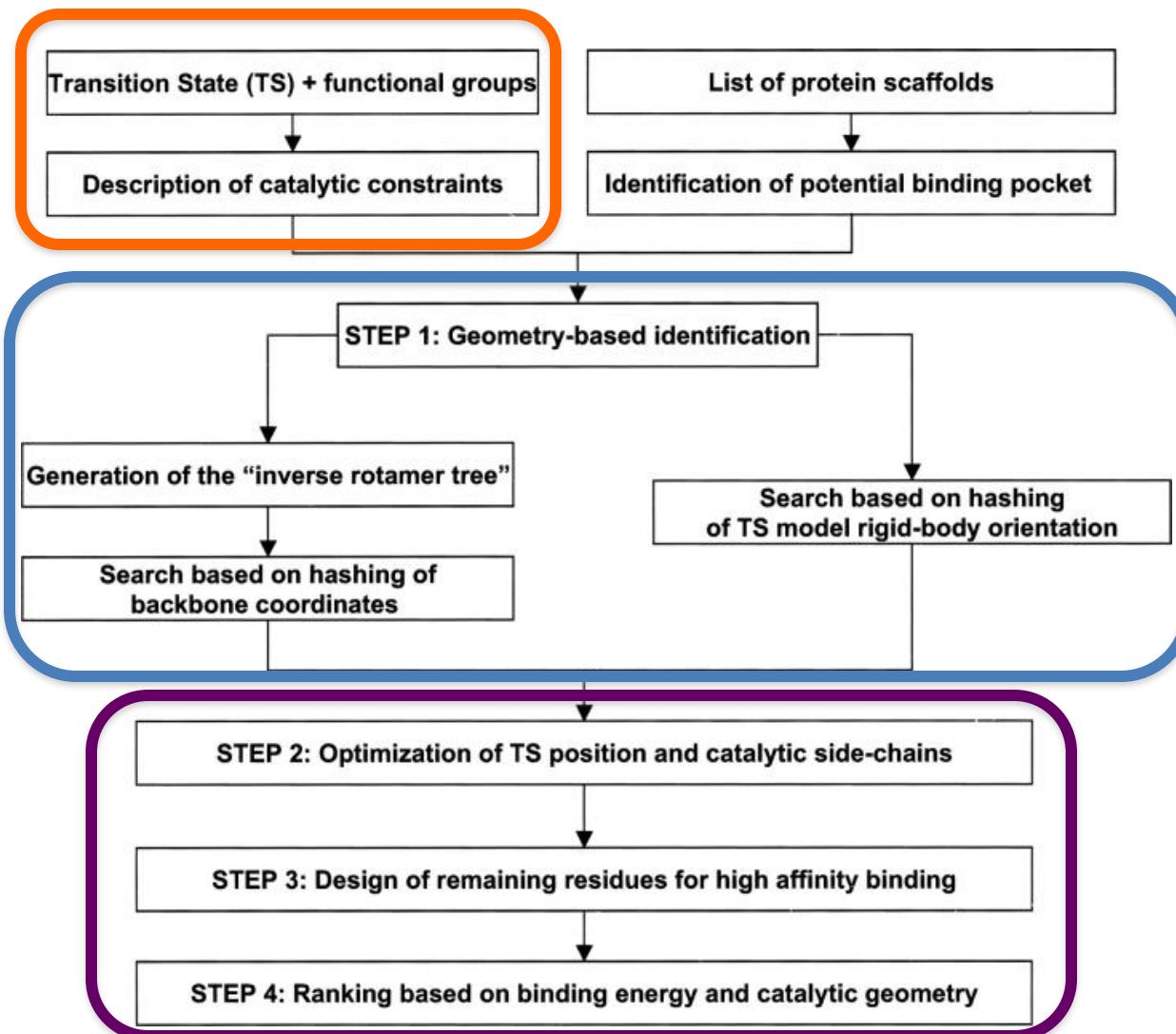
# Design of a novel enzyme

*Roethlisberger et al. 2008; Liang et al., 2008*



# Design of a novel enzyme

*Roethlisberger et al. 2008; Liang et al., 2008*



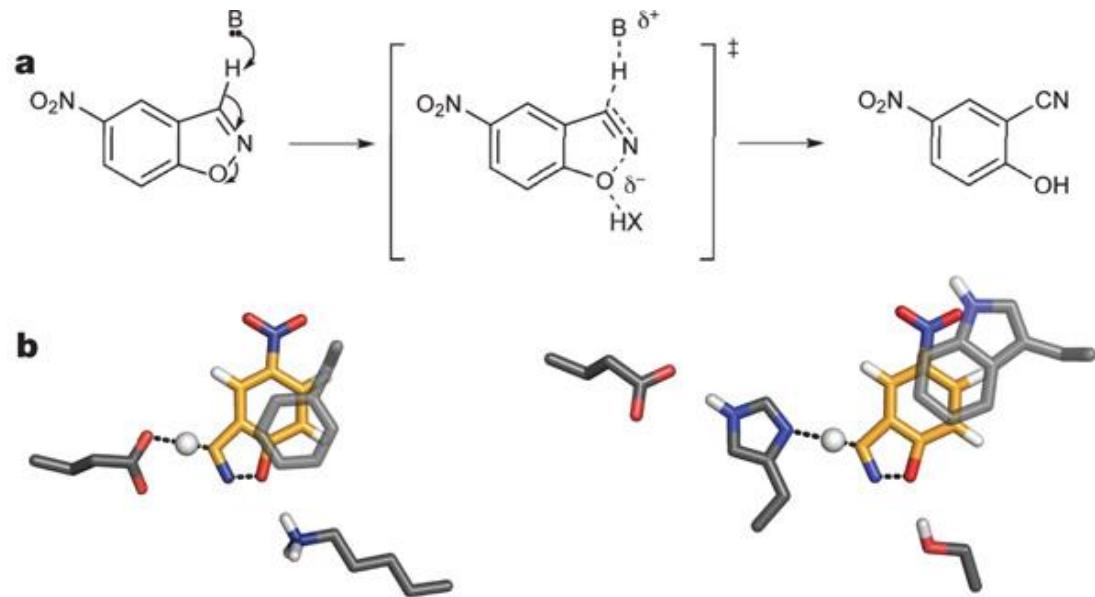
Approach:

1. Model transition state of reaction (QM)
2. Stabilize with carefully placed chemical groups around it
3. Graft resulting active site into an existing protein
4. Alter the sequence of the protein to accommodate the active site

# Model transition state

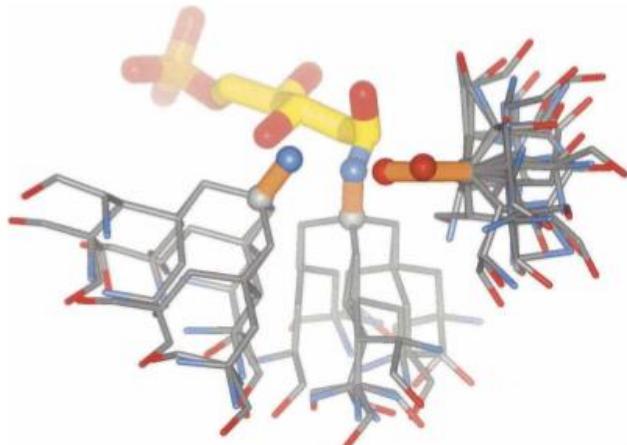
- Kemp Elimination
- Water mediated

Model transition state

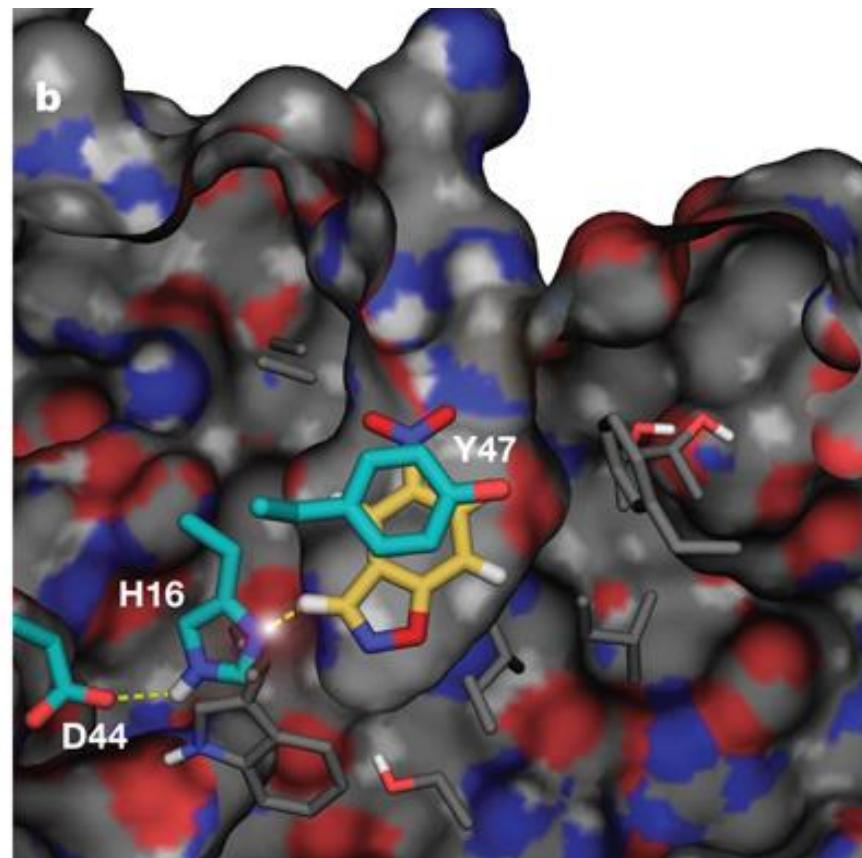
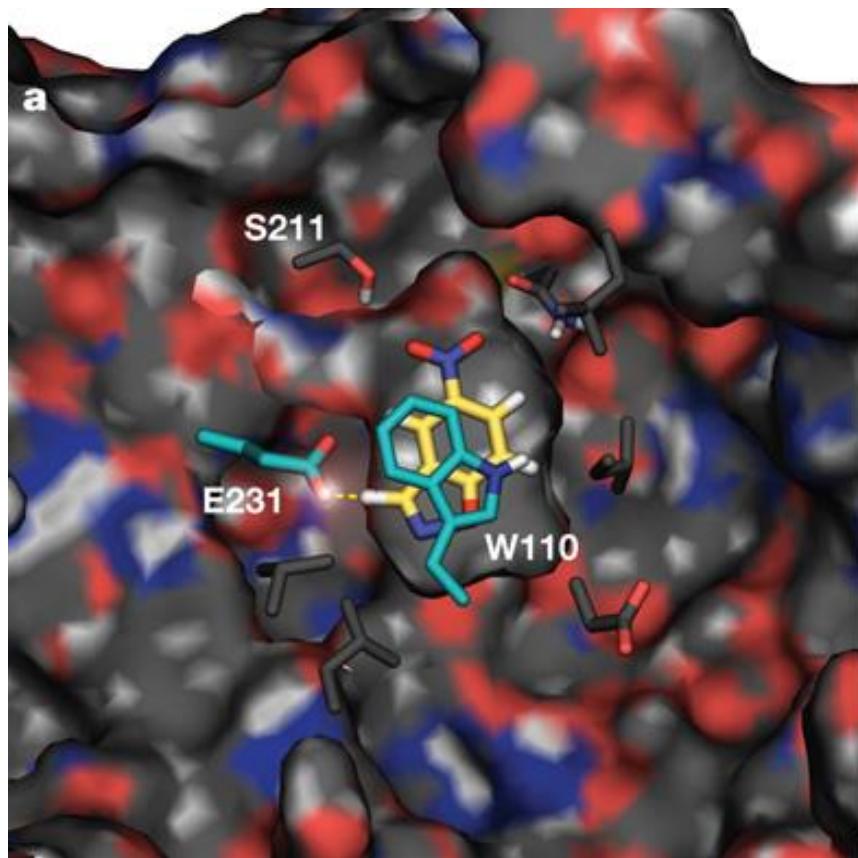


# Search for Template

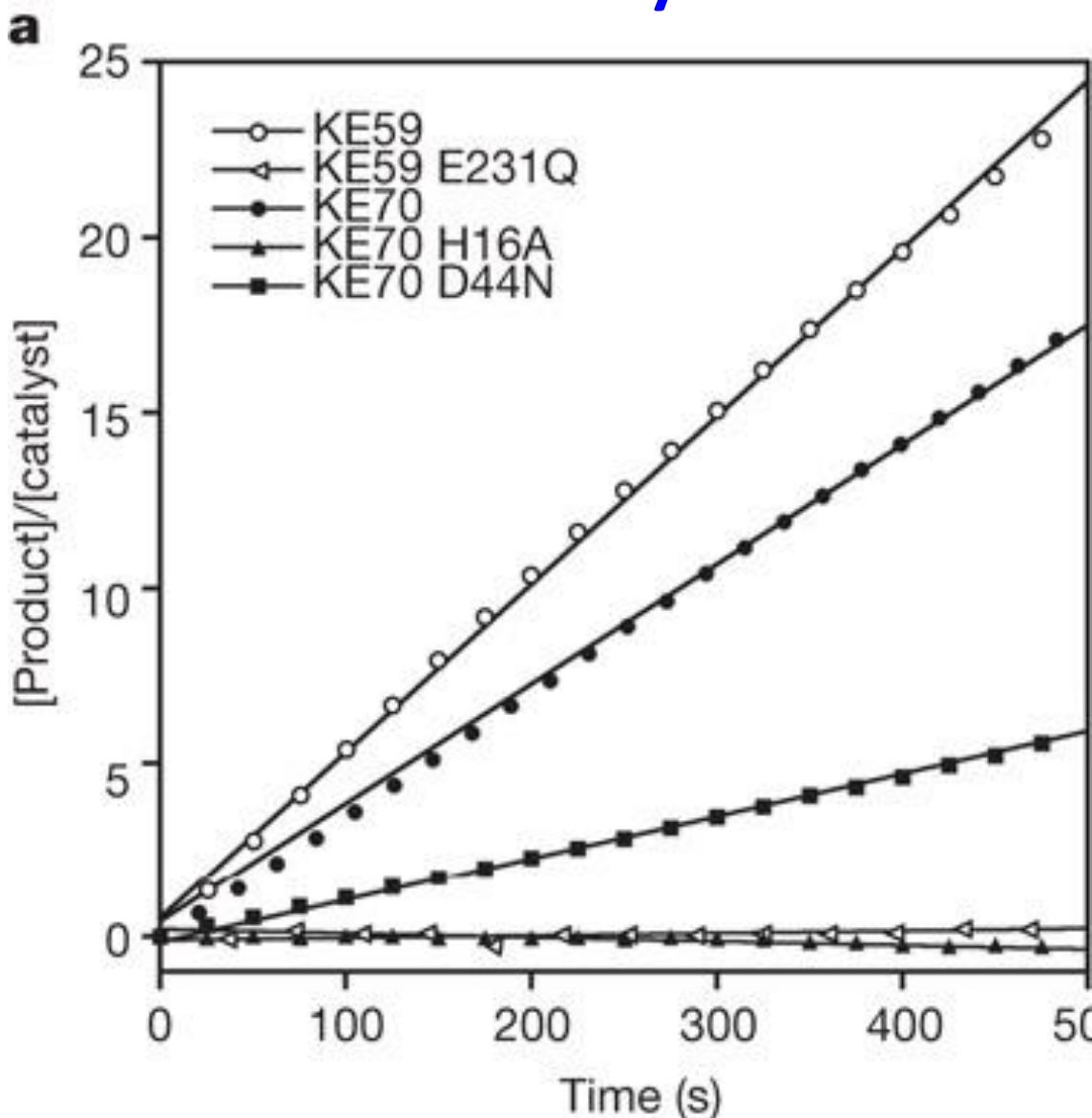
- Inside-out:
  - Build inverse rotamer tree starting from catalytic site
  - Search for fitting backbone templates (geometric hashing)
- RosettaMatch: Outside-in:
  - Place side chains and transition state model at each position
  - Search for transition state model orientations that fit several positions



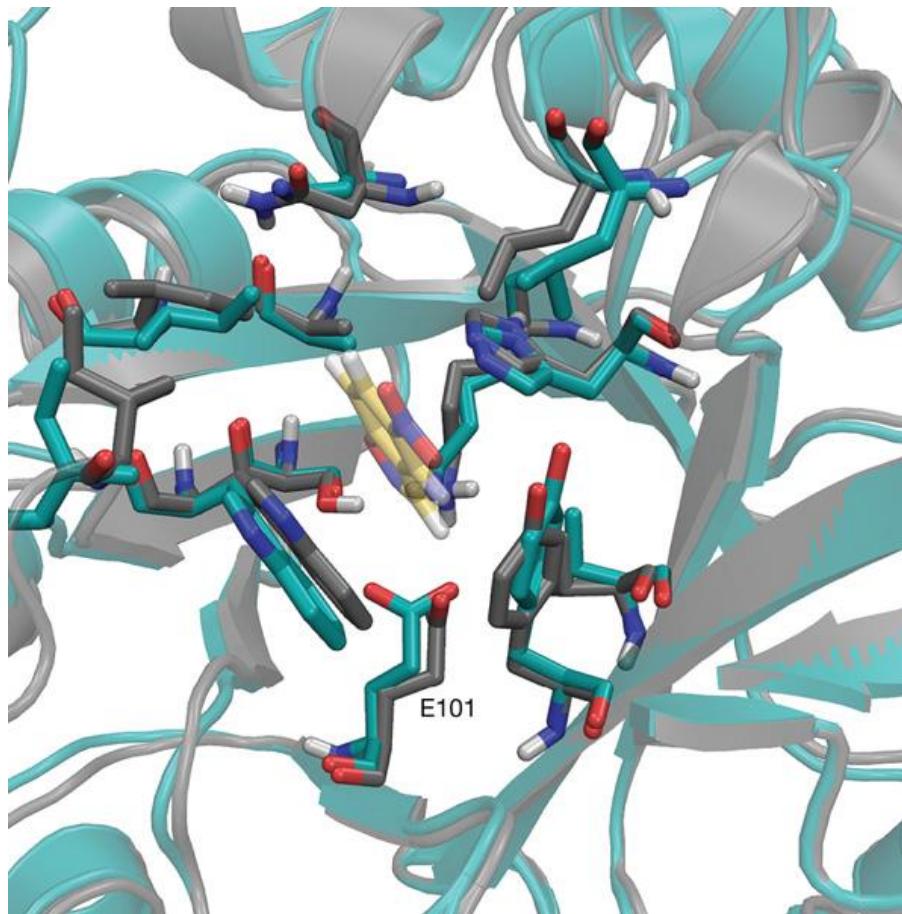
# Find match



# Validation 1: enzyme is active



# Validation 2: accurate structure prediction



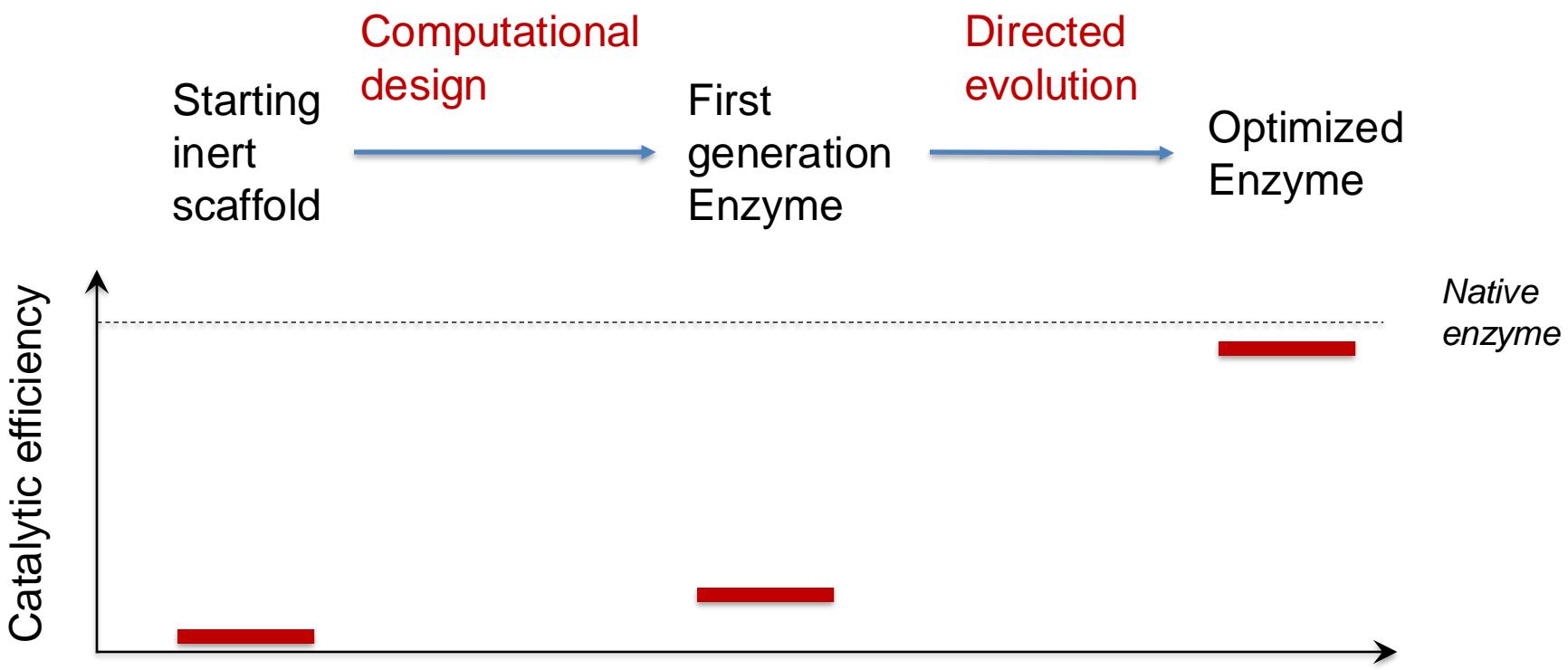
Is this all?

Is the enzyme design problem  
solved?

Designed HG3:  $k_{cat} \sim 1 \text{ s}^{-1}$

Native kemp eliminases:  $k_{cat} \sim 400-500 \text{ s}^{-1}$

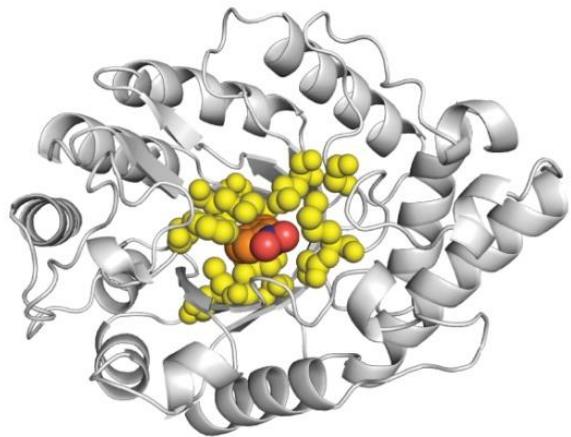
# Precision is essential for efficient catalysis in an evolved Kemp eliminase (Blomberg, Nature 2014)



# Directed evolution of Kemp eliminase HG3 (Blomberg, Nature 2014)

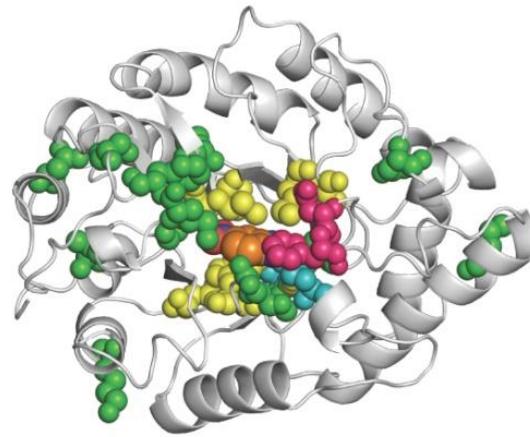
Computational design: HG3

**a**

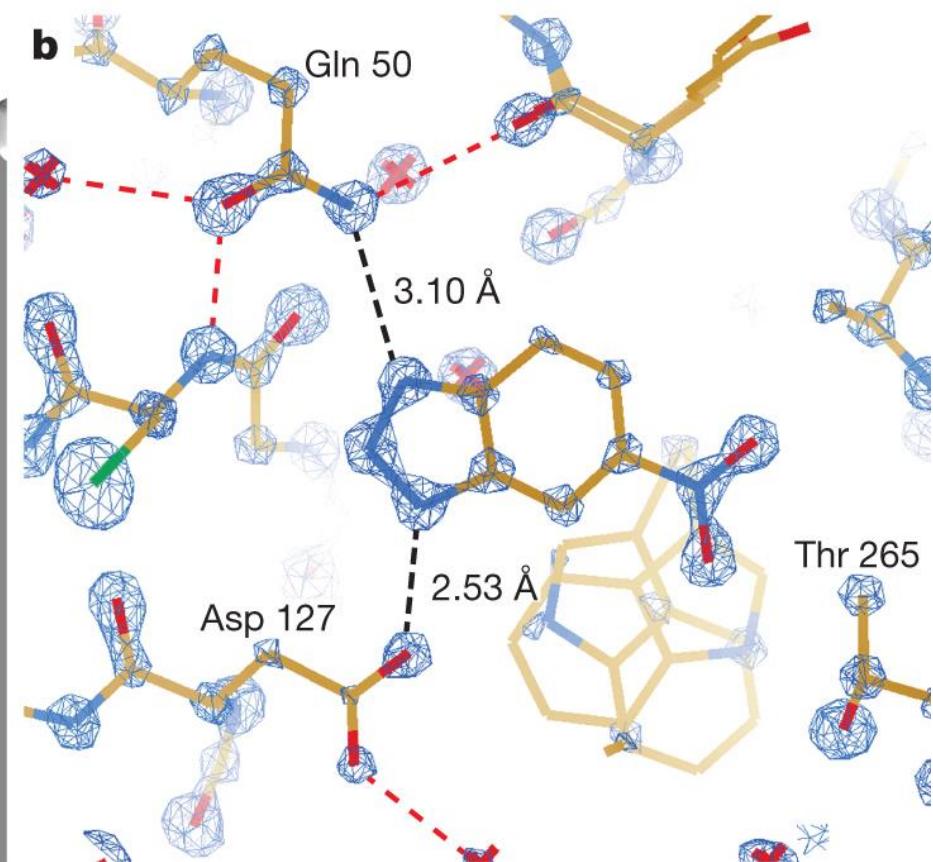
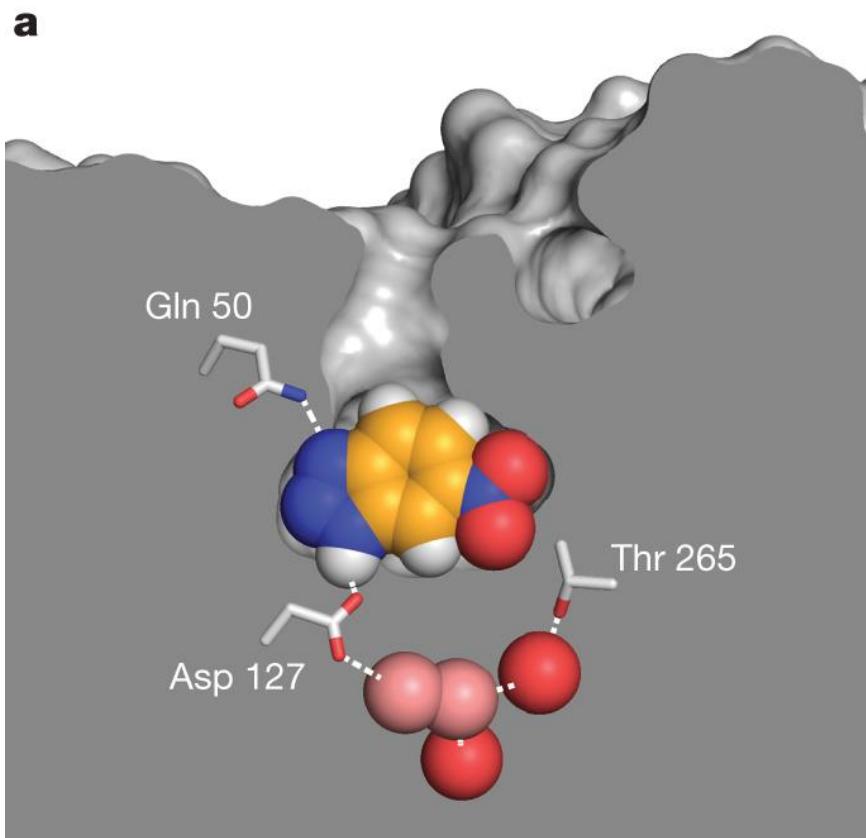


Directed evolution: HG3.17

**b**



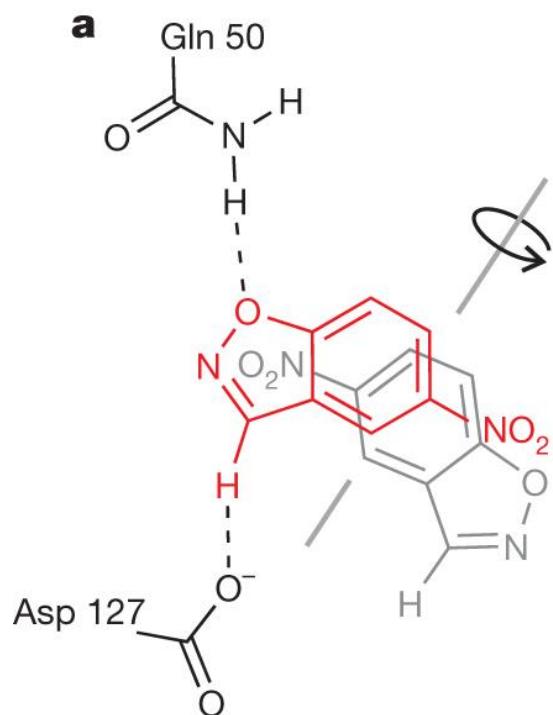
# Crystal structure of HG3.17 (Blomberg, Nature 2014)



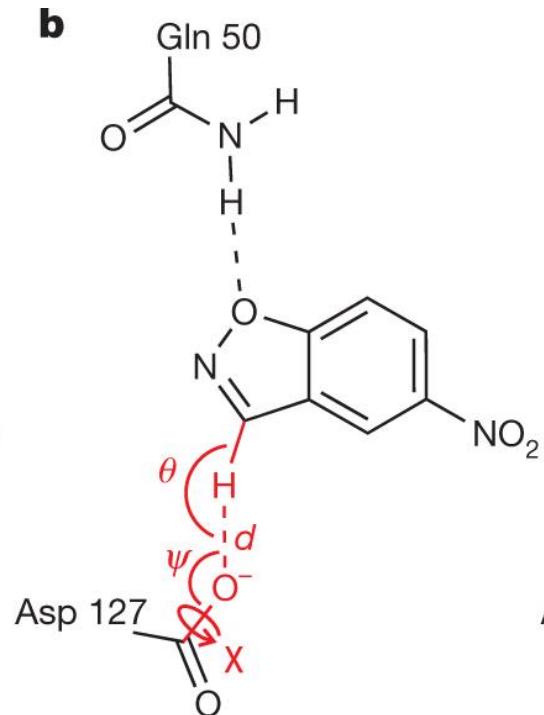
# Catalytic improvement of HG3

(Blomberg, Nature 2014)

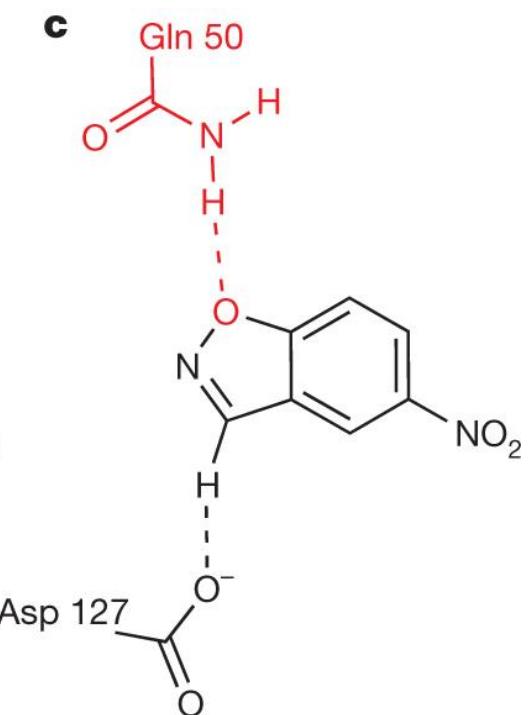
Elimination of a  
on unproductive  
binding mode



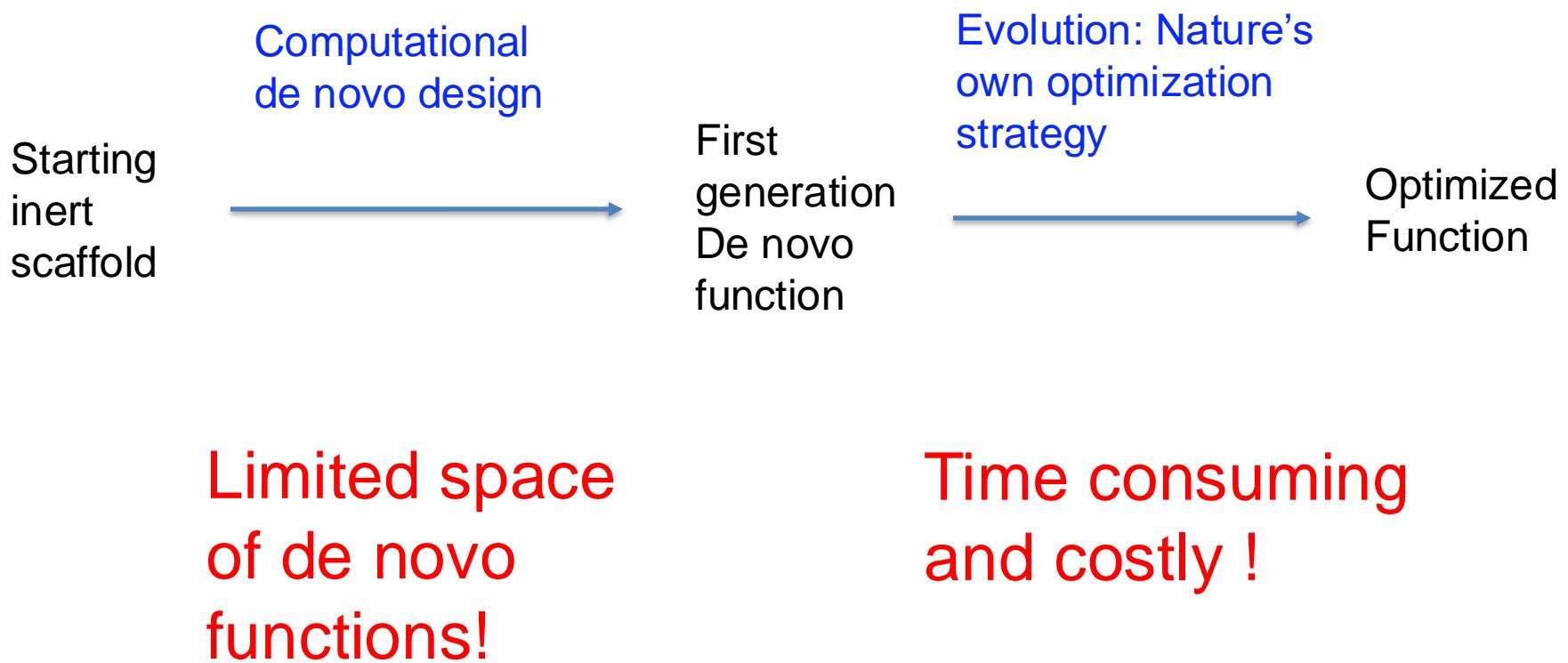
Efficient proton  
transfer



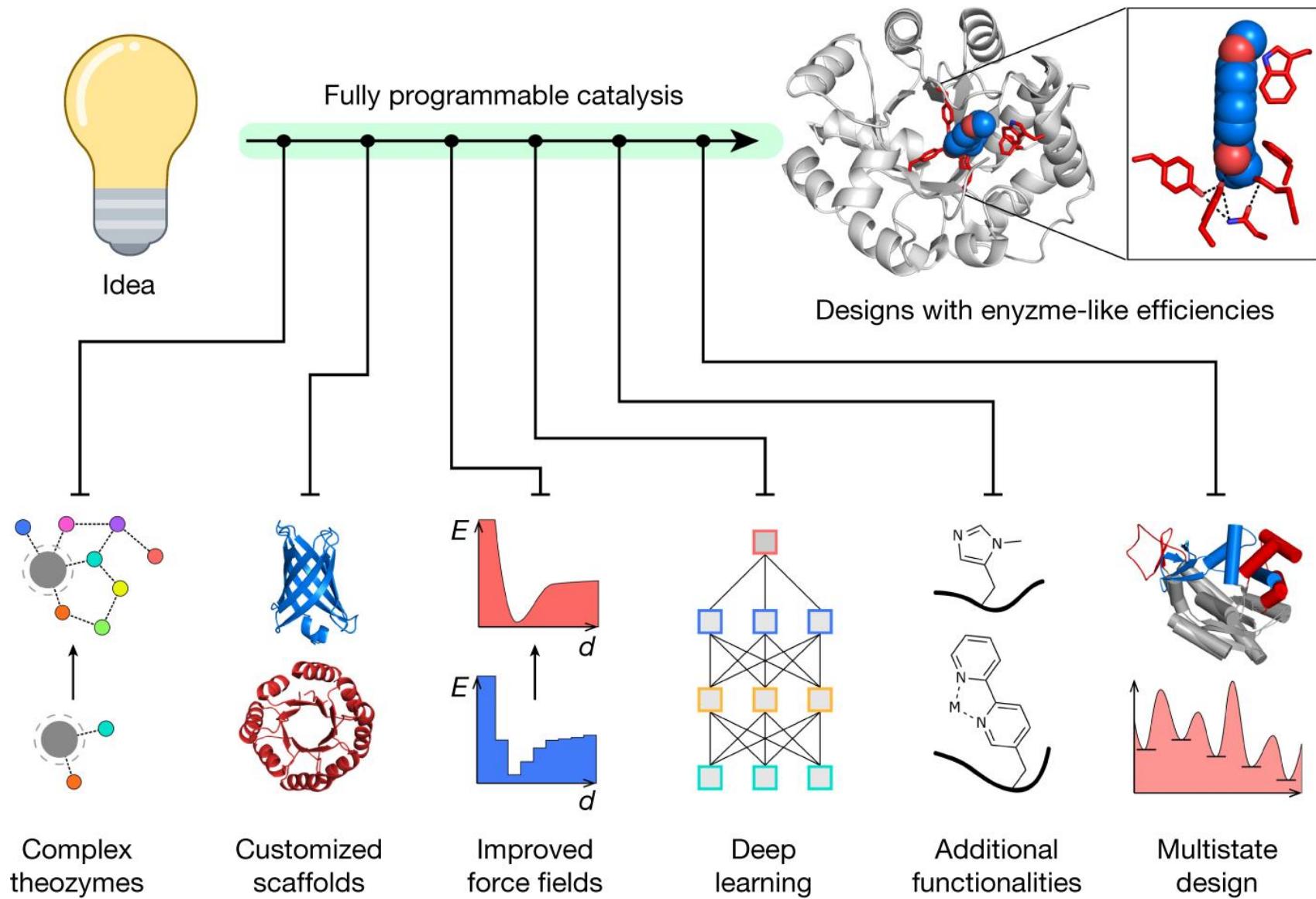
Optimal TS  
stabilization



# Take home messages

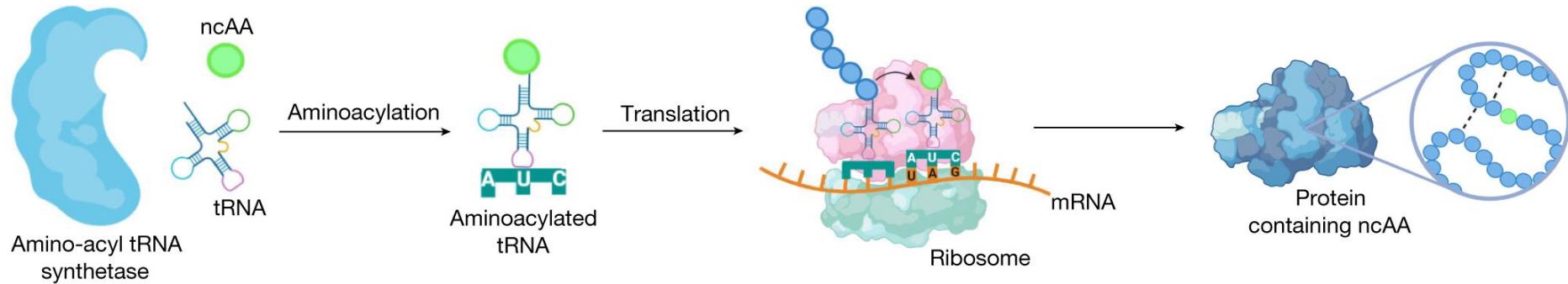


# A road map to better enzymes



# One future direction: non canonical AA for expanded chemical space

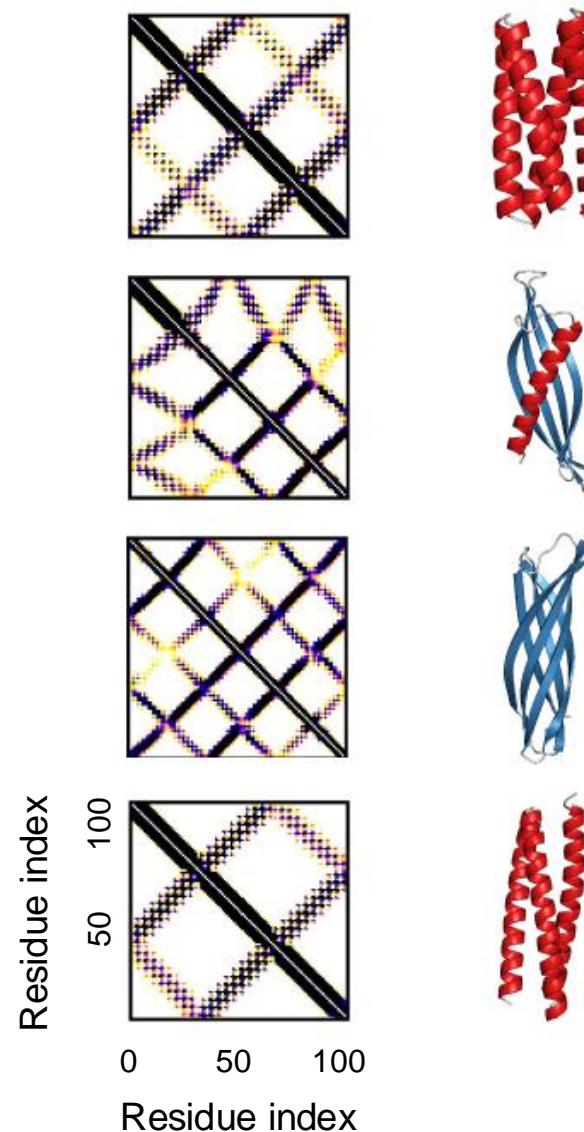
a



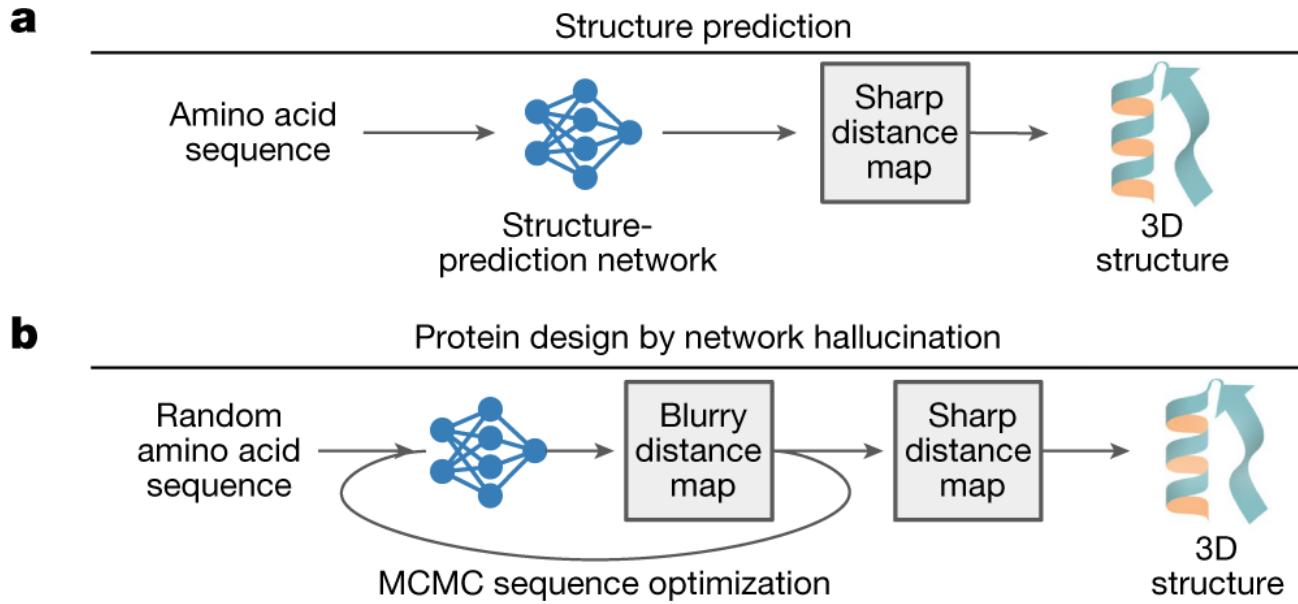
# De novo design of luciferases using deep learning

Each protein structure is characterized by a matrix of residue-residue contacts

Neural networks learn these contact patterns



# De novo design of luciferases using deep learning



# De novo design of luciferases using deep learning

**b**

Protein design by network hallucination

Random  
amino acid  
sequence



Blurry  
distance  
map

Sharp  
distance  
map

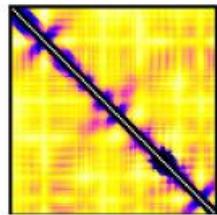


3D  
structure

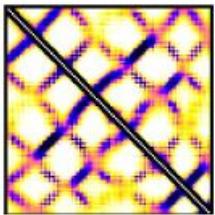
MCMC sequence optimization

MCMC step

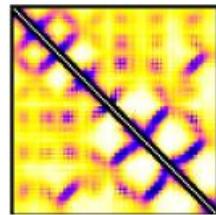
0



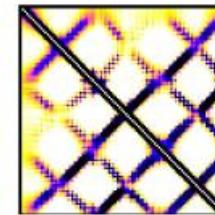
1000



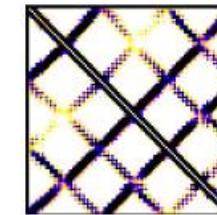
5000



10000



40000



Residue index

0

50

100

0 50 100  
Residue index

Residue index

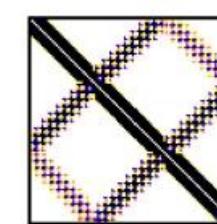
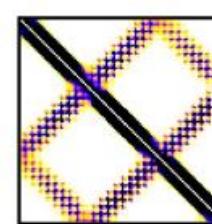
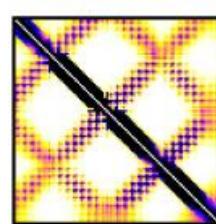
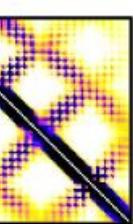
0 1  
Predicted probability of  
 $C_{\beta}-C_{\beta}$  distance  $< 10 \text{ \AA}$

Residue index

0

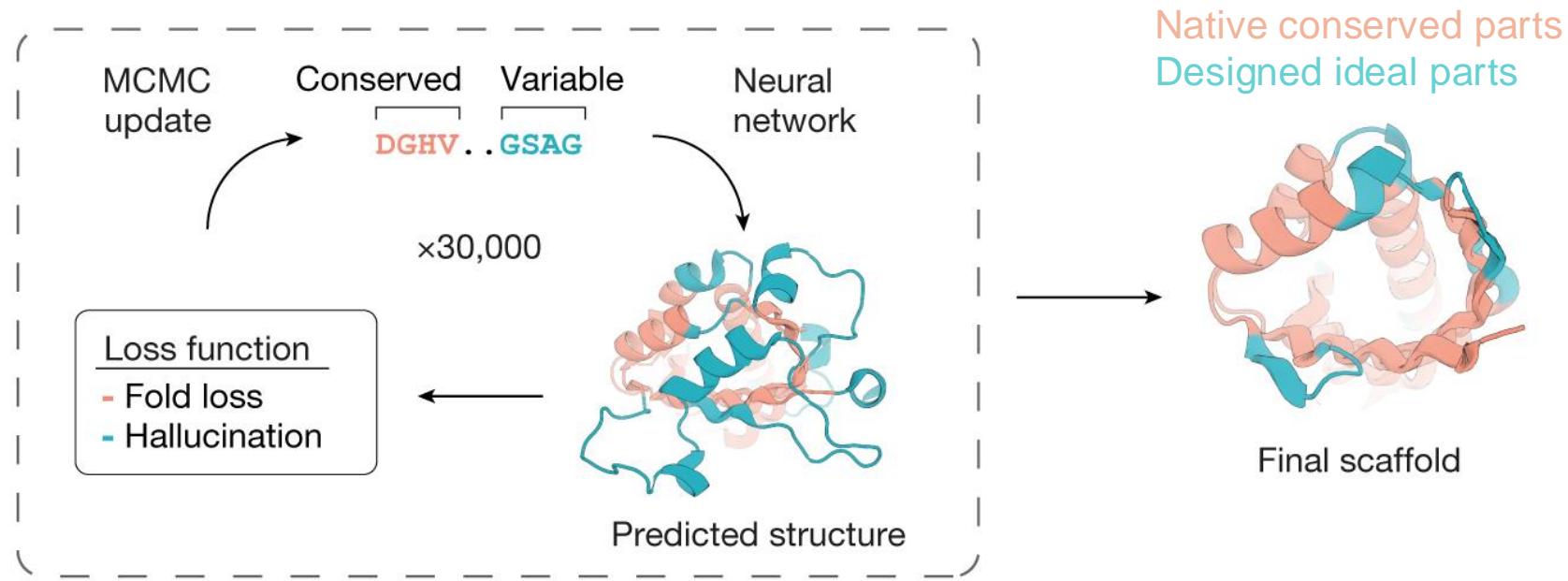
50

100

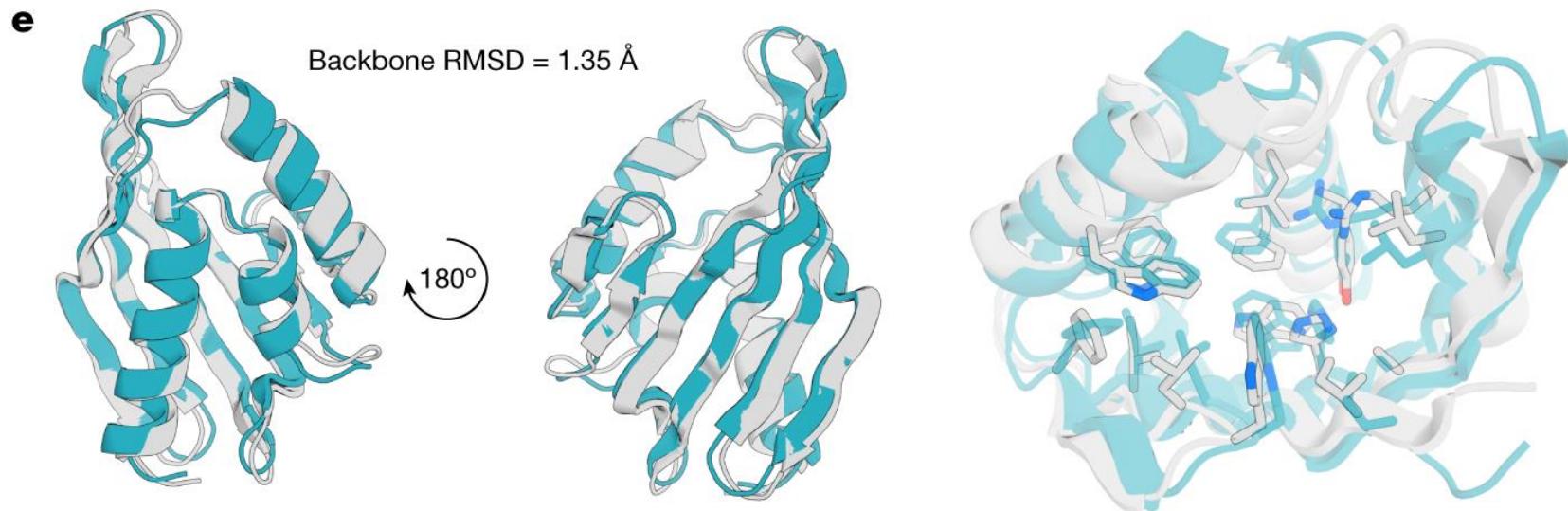
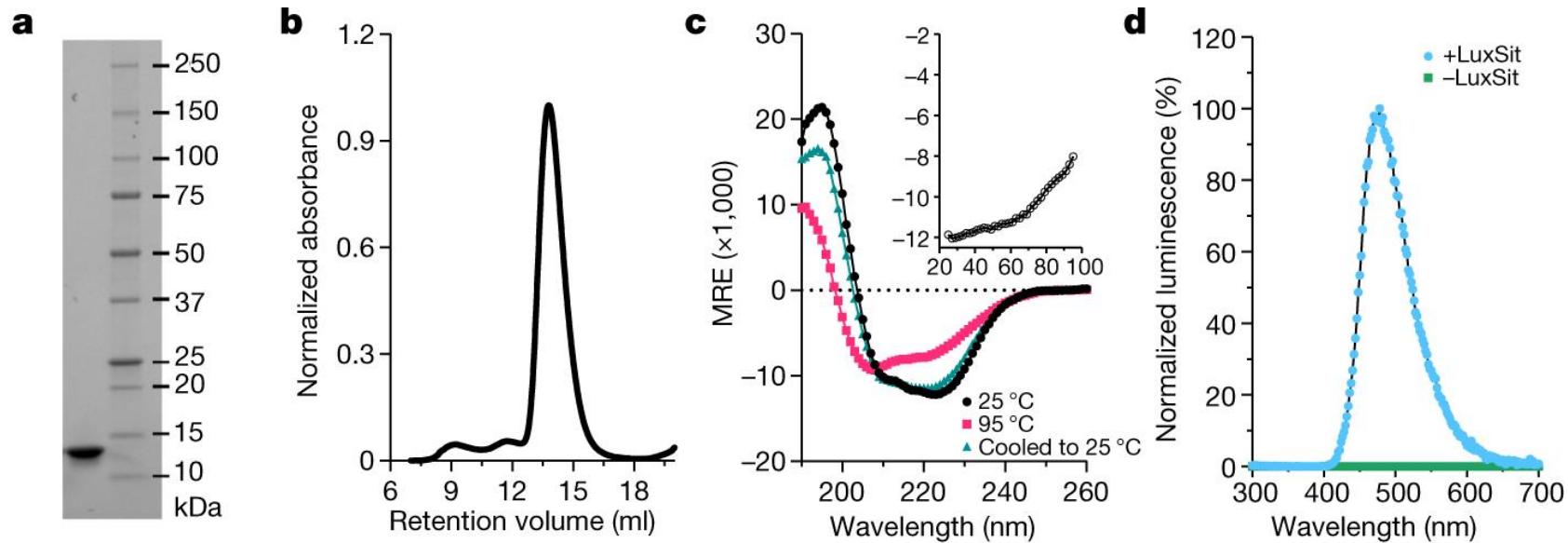


# De novo design of luciferases using deep learning

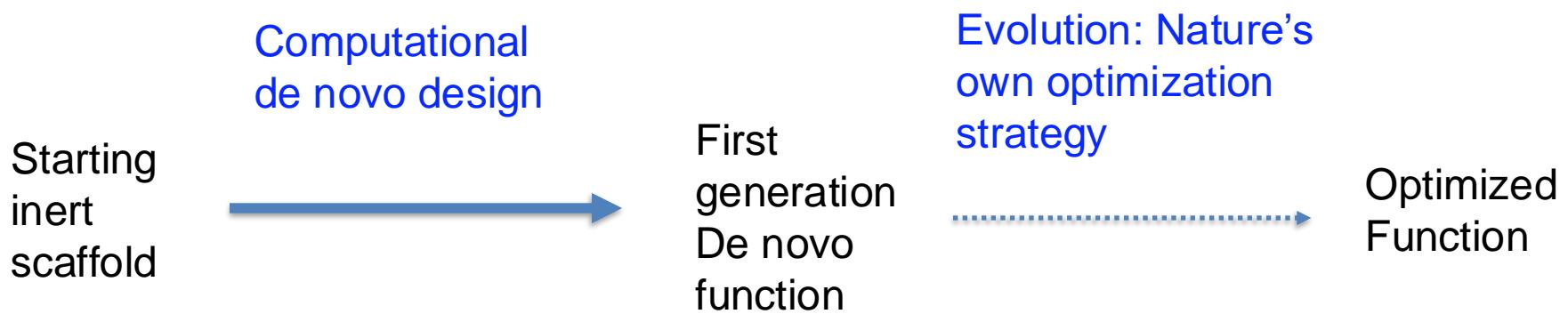
a



# De novo design of luciferases using deep learning



# Take home messages



Deep learning

Artificial chemistry

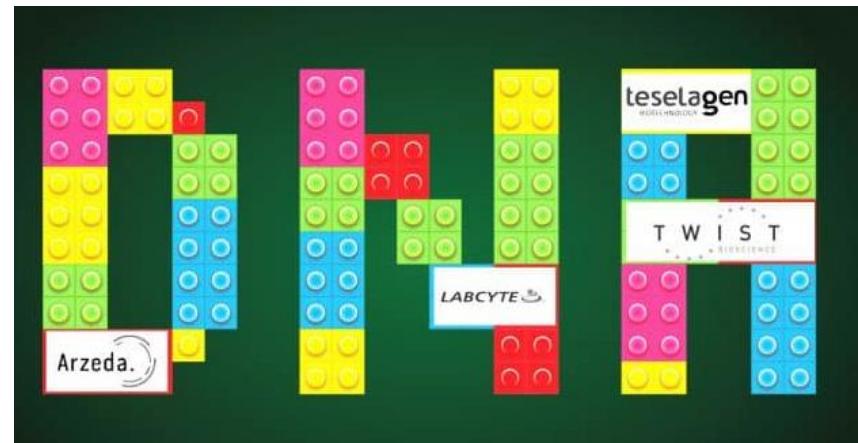
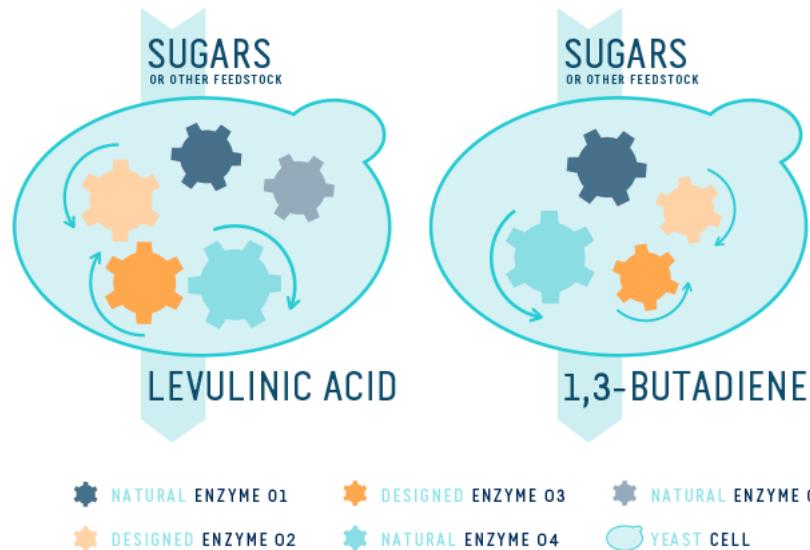
Many others

# Enzyme Design for a more sustainable world



Arzeda is harnessing the power of computational protein design to build novel enzymes and discover new pathways that enable cost effective, sustainable production of value-added specialty chemicals and ingredients

## 02 ARZEDA'S DESIGNER CELL FACTORY



# Protein Design – Examples overview

1. De novo protein functional fold

2. Enzyme

3. Ligand biosensor design

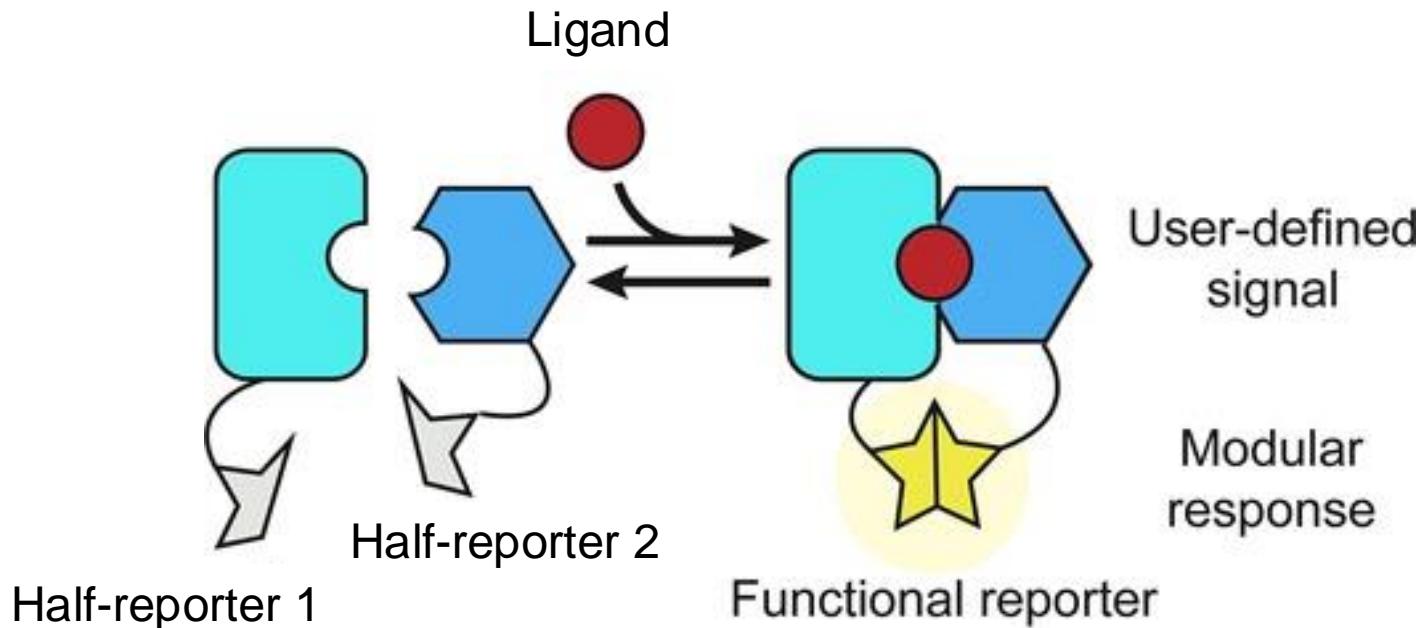
# Ligand biosensor design

What is a biosensor?

What can it be useful for?

Why do we care?

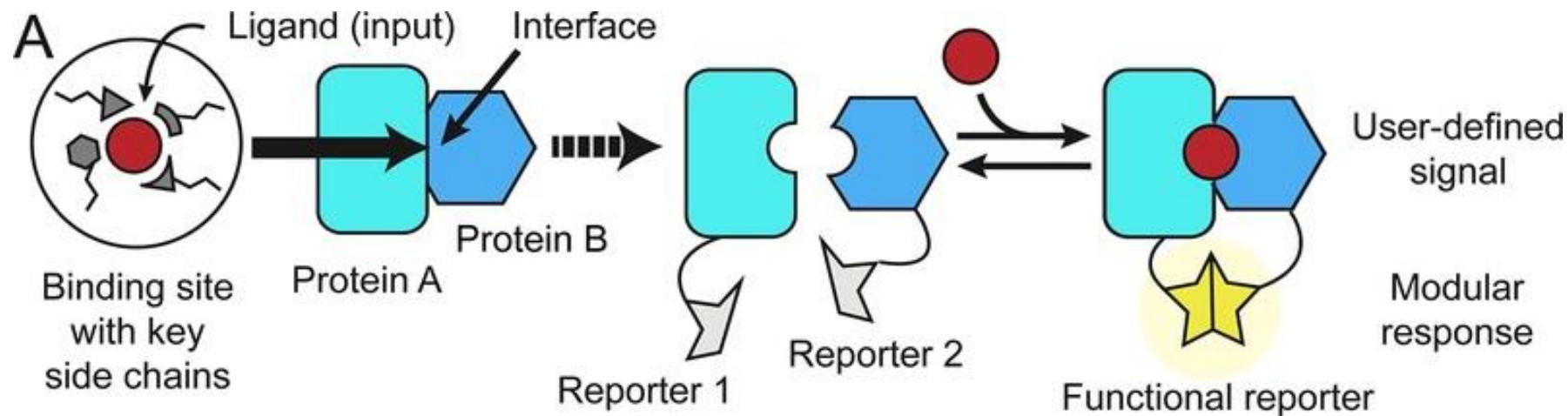
# Ligand biosensor design



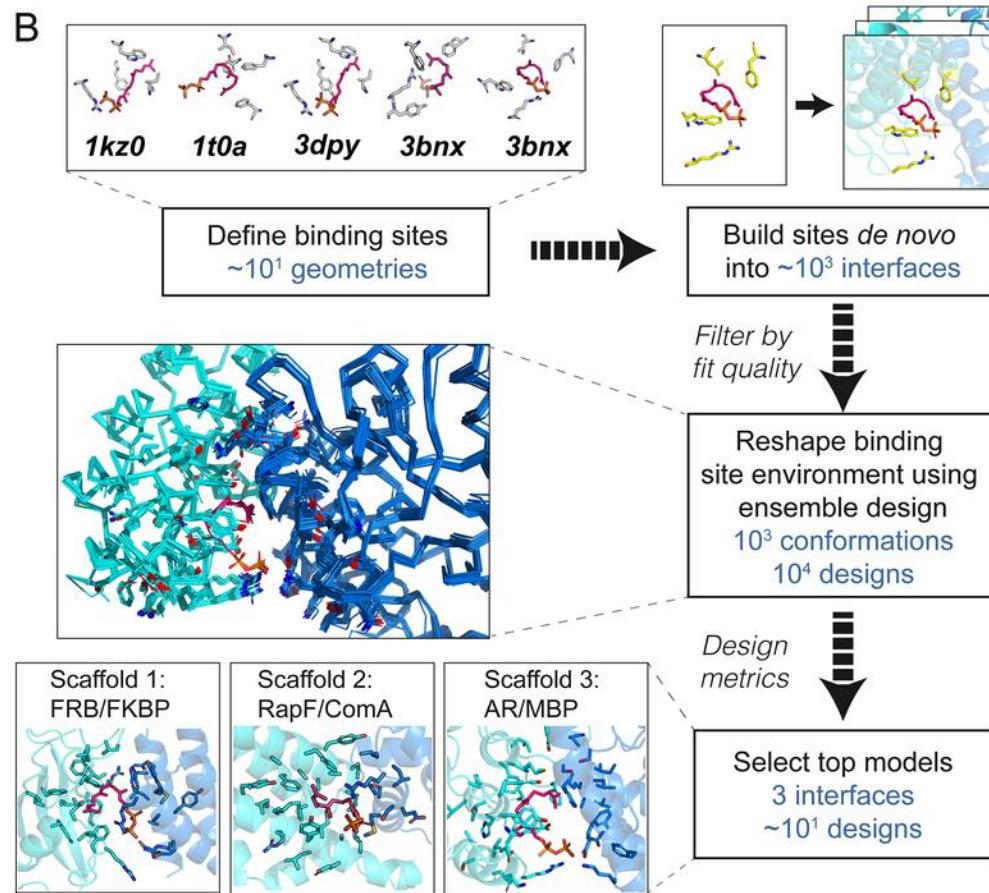
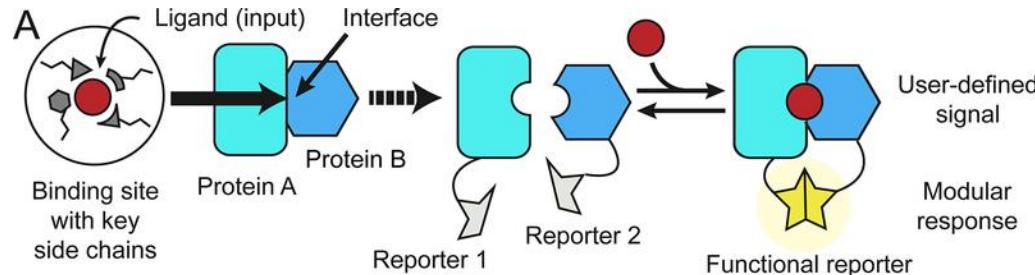
A wide range of applications

Diagnostics  
Detection of contaminants  
Data storage

# Ligand biosensor design

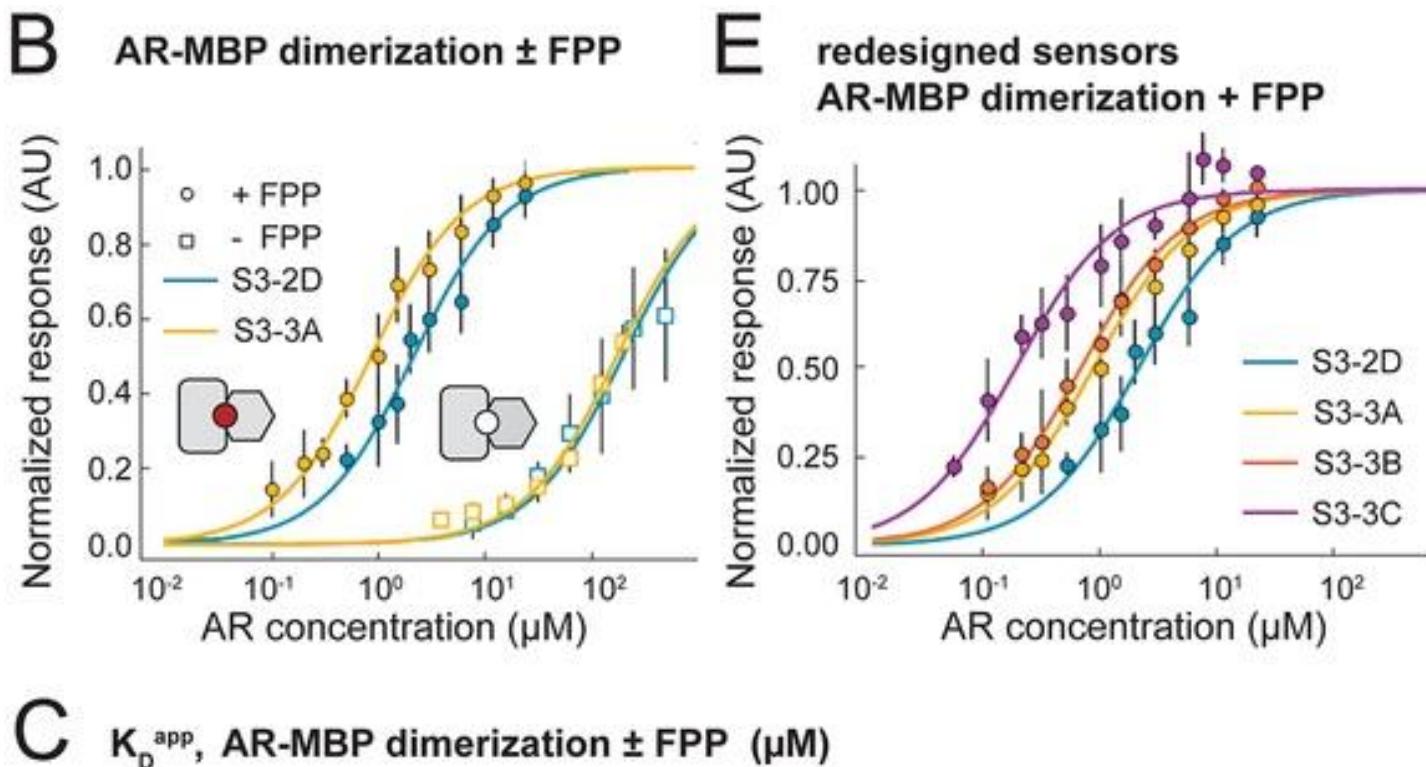


# Ligand biosensor design



(Glasgow 2020)

# Ligand biosensor design



**C**  $K_D^{\text{app}}$ , AR-MBP dimerization  $\pm$  FPP ( $\mu\text{M}$ )

	S3-2D	S3-3A	S3-3B	S3-3C
+FPP	$2.1 \pm 0.18$	$0.87 \pm 0.06$	$0.67 \pm 0.03$	$0.17 \pm 0.02$
-FPP	$>200^{\#}$	$>200^{\#}$	$14.0 \pm 1.09$	$6.16 \pm 0.31$
Fold change	>100	>200	20.8	36.2

# low estimate

# Ligand biosensor design

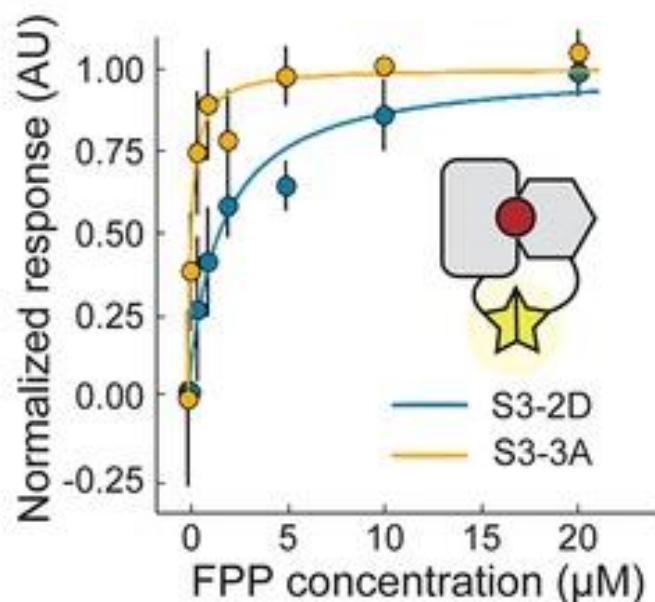
D  $K_D$ , FPP binding to AR or MBP (μM)

S3-2D AR	$6.1 \pm 1.4$
S3-2D MBP	$>1000^{\#}$

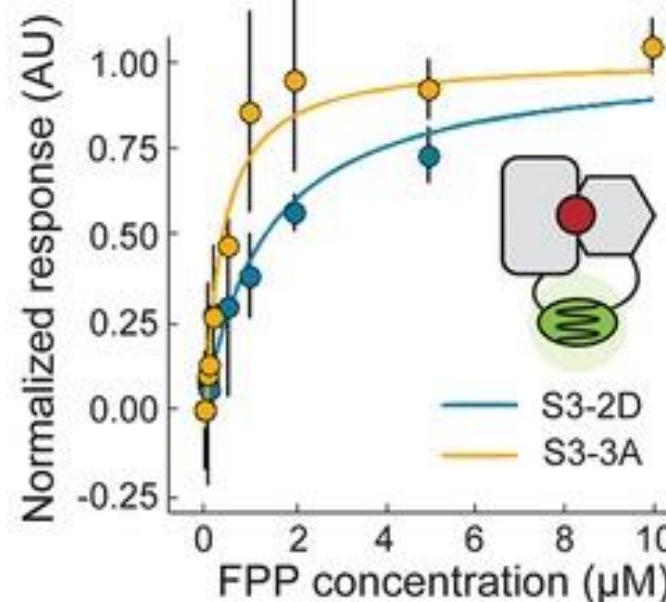
F  $K_D^{app}$ , FPP sensitivity of sensors (μM)

	S3-2D	S3-3A
Luminescence	$1.6 \pm 0.47$	$0.18 \pm 0.05$
Fluorescence	$1.4 \pm 0.50$	$0.33 \pm 0.13$

G Luminescence reporter

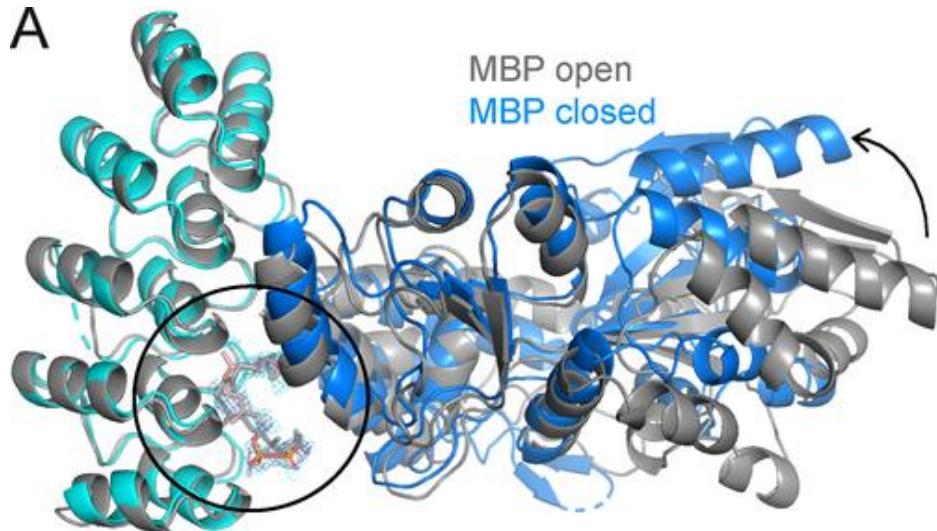


H Fluorescence reporter

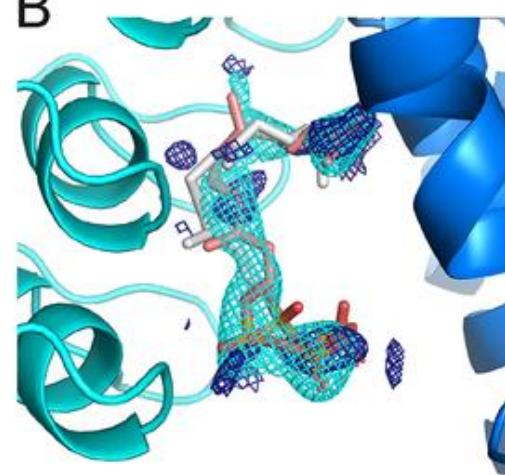


# Ligand biosensor design

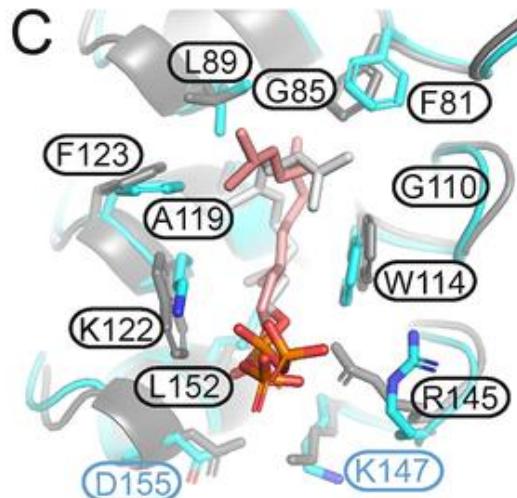
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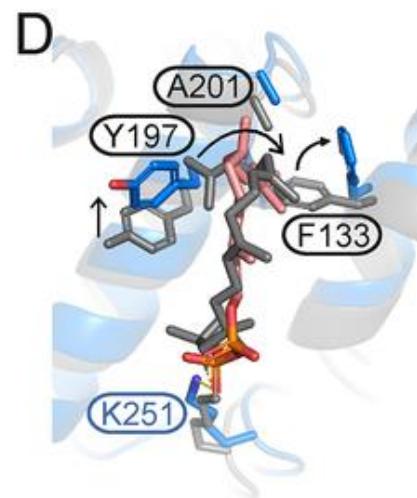
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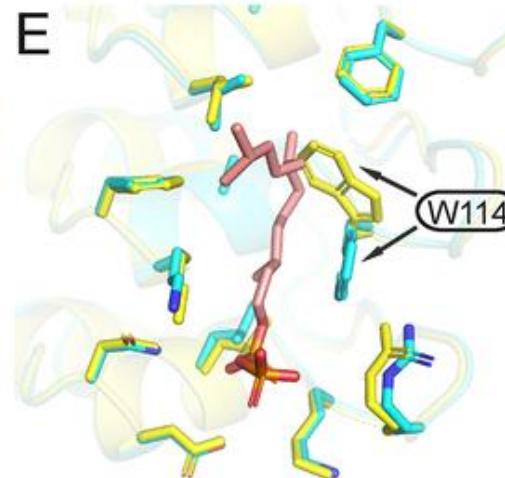
C



D



E



S3-2D  
design model

S3-2D AR  
crystal structure

S3-2D MBP  
crystal structure

S3-2D FPP  
crystal structure

S3-2D Apo  
crystal structure

# Ligand biosensor design

Have they solved the biosensor design problem?

Where do you see key limitations?

What could be the next steps?

# Take home messages

1. Protein binding vs folding differences (interactions, conformational space)
2. Solutions for binding site design (starting from precise ligand conformation)
3. Computational design solutions are often suboptimal. Need time-consuming optimization by directed evolution
4. AI-based methods are emerging that can create protein scaffolds stabilizing precise ligand structures